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March 11, 2015

Wess Safford, Air Quality Engineer South West Clean Air Agency 11815 NE 99<sup>th</sup> Street, Suite 1294 Vancouver, WA 98682

Re: 2015 Kiln Emissions Testing

Enclosed is the 2015 kiln emission testing results conducted by Michael Milota at OSU on February 1, 2015.

If you have any additional questions or comments, please feel free to contact me.

Thank You,

Jason Thompson Safety / Environmental Manager 360-748-0178 thompsonjr@chwa.com



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Randy
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Traci
Tina
File

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Total hydrocarbon and HAP emissions from the drying of red alder lumber

> Report to Cascade Hardwood LLC 158 Ribelin Road, Chehalis, WA 98532 Phone: 360-748-0178 Fax: 360-740-5118 Contact: Jason Thompson Air Discharge Permit #04-2566R2

> > Report by

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March 5, 2015



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March, 2015

## Summary

One charge of red alder 5/4 lumber was dried in a small kiln at Oregon State University. The kiln dry- and wet-bulb temperatures were based on a schedule provided by Cascade Hardwoods. The maximum temperature was 170°F (77°C). The air velocity was 750 feet per minute (3.7 m/s). The kiln was indirectly heated with steam. The amount of air entering the kiln was regulated to control humidity.

A JUM VE-7 total hydrocarbon analyzer was used to measure organic emissions following EPA Method 25A. The results are shown in Table 1.

**Table 1.** Summary of total hydrocarbon results to 8% moisture content. VOC units are pounds per thousand board feet as carbon.

Charge	Initial MC	Final MC	Time to 8%	VOCB
	%	%	hr:min	lb/mbf
Red alder	106.1	8.0 <sup>A</sup>	89:28	0.284

A actual time to 6.7% MC was 106:53 hours

<sup>B</sup> VOCs are reported at 8% moisture content

NCASI Method ISS/FP-A105.01 was used to measure the MACT HAP emissions. The results are shown in Table 2. The sum of the HAPs emitted was 0.127 lb/mbf for the red alder.

Table 2.Summary of HAP results for moisture content and time shown in Table1. Emissions units are pounds per thousand board feet.

	Methanol	Phenol	Form- aldehyde	Acet- aldehyde	Propion- aldehyde	Acrolein
Red alder	0.062	0.0	0.0007	0.0623	0.0005	0.0007

<sup>A</sup> None detected

#### 1. Description of source

The tested source is a lumber dry kiln. Lumber destined for the mill's kiln was sampled and tested in a small-scale kiln at Oregon State University.

Mill personnel reported that the harvest location was Southwest Washington and Northern Oregon. The logs were stored in the logyard for 10 days on pavement without sprinkling. The logs were sawed on February 5, 2015, wrapped, and loaded to be delivered to OSU on February 5, 2015.

Enough 4-foot pieces of wood for three charges of lumber were delivered to Oregon State by an employee of Cascade Hardwoods. The wood was palletized and wrapped in plastic at the mill to prevent moisture loss during transport. The wood appeared to be fresh.

At OSU, the 4-foot pieces were sorted February 5. One third was randomly pulled from the pile and used for the charge. It was wrapped and stored in a cooler at 40°F. The remaining wood was wrapped in plastic and placed in a freezer at 10°F.

# 2. Date and time of test

The charge was dried from February 9 at 6 am until 5 pm February 13, 2015. Drying was done under the supervision of Mike Milota at Oregon State University. Students were used to monitor parts of the test.

# 3. Results

#### Total hydrocarbon

See Table 1, page 1, for a summary of the hydrocarbon results. Details for each sampling interval are tabulated (Table 3) and the hydrocarbon emissions are summarized graphically (Figures 1-3) here. All emission data is presented in detail in electronic form in Appendix 2.

An interval is the period between analyzer calibrations, about six hours of data. The interval time periods shown in the table include the calibration times and mass calculations are adjusted to account for these. Sampling occurred for approximately 95% of the drying time.

Figure 1 shows total hydrocarbon concentration (left scale) and dry gas vent rate (right scale) versus time. Concentration has a large peak at 12.6 hours during the period when venting is low. Concentration is reduced by 24 hours due to a much higher vent rate. In general, the concentration then increases though the schedule as venting decreases. The vent rate is quite variable in the beginning of the

schedule due to heating of the wood and a high rate of moisture release from the wood.

Figure 2 shows the cumulative hydrocarbon emissions (left scale, smooth line) and the rate of emissions (right scale, jagged line) versus time. The cumulative value is the emissions up to any point in time in the schedule. The rate is how much is coming out per unit time. The maximum emission rates occurred between 12 and 36 hours, after which it steadily decreases as the moisture loss from the wood slows. Spikes in the lines correspond to schedule changes.

Figure 3 shows the total hydrocarbon emissions as a function of wood moisture content. This graph would be useful for predicting emissions at various final moisture content levels. As is typical, the hydrocarbon release rate is linear with moisture loss at the lower moisture content.

Sample	Time	Average	Flow	rate	THC mass	THC con	centration	THC rate	THC mass	THC rate	1	Average	
Run	hrs	Humidity kg/kg	Dry @68 I/min	Wet @68 I/min	as C g	wet ppmv	dry ppmv	as C g/hr	as C Ibs/mbf	as C lb/hr/mbf	Wood MC %	Air MC %	Anal. MC %
1	3.51	0.041	35.8	38.2	0.04	3.0	3.1	0.01	0.001	0.0003	106.0	6.2	6.2
2	5.06	0.071	156.8	174.8	0.57	7.2	8.1	0.11	0.014	0.0028	103.2	10.3	10.3
3	3.56	0.121	45.1	53.9	0.48	36.1	44.5	0.14	0.012	0.0033	98,9	16.3	16.3
4	5.96	0.180	57.1	73.6	1.30	35.7	46.1	0.22	0.032	0.0053	95.6	22.4	13.5
5	5.21	0.170	108.3	137.9	1.24	21.3	27.3	0.24	0.030	0.0058	88.8	21.5	12.9
6	6.71	0.153	189.0	235.6	1.66	11.7	14.5	0.25	0.041	0.0060	73.8	19.8	11.9
7	1.05	0.151	189.4	235.3	0.25	11.2	13.9	0.24	0.006	0.0058	63.1	19.5	11.7
8	5.01	0.151	173.7	215.9	1.24	12.8	15.9	0.25	0.030	0.0061	55.1	19.5	11.9
9	6.06	0.150	149.4	185.5	1.31	13.0	16.2	0.22	0.032	0.0053	42.4	19.5	11.9
10	5.81	0.150	99.1	123.0	0.93	14.5	18.0	0.16	0.023	0.0039	31.5	19.5	11.9
11	5.91	0.150	66.0	81.9	0.68	15.7	19.4	0.11	0.017	0.0028	24.9	19.4	11.8
12	8.11	0.149	55.1	68.4	0.73	15.0	18.6	0.09	0.018	0.0022	18.5	19.4	12.2
13	4.01	0.149	25.5	31.6	0.22	19.4	24.0	0.05	0.005	0.0013	14.9	19.4	13.0
14	5.96	0.149	25.5	31.6	0.33	19.5	24.2	0.06	0.008	0.0014	13.1	19.4	13.0
15	6.01	0.148	16,7	20.7	0.23	20.2	25.1	0.04	0.006	0.0009	11.3	19.3	12.9
16	6.16	0.148	11.5	14.2	0.16	20.0	24.8	0.03	0.004	0.0006	10.1	19.3	13.2
17	5.41	0.147	21.6	26.7	0.23	18.3	22.6	0.04	0.006	0.0010	8.7	19.1	13.1
Sum	89.47				11.6				0.284				
Average		0.140	83.9	102.9		17.3	21.6	0.13		0.0032	La L		1

Table 3. Summary of results for each sampling interval for total hydrocarbon.



Figure 1. Hydrocarbon concentration and vent rate versus time 1.



Figure 2. Cumulative hydrocarbon emissions (left scale, smooth line) and the rate of emissions (right scale, jagged line) versus time.



Figure 3. Total hydrocarbon emissions as a function of wood moisture content.

# HAPs

See Table 2, page 1, for a summary of the HAP results. Details for each sampling interval are tabulated (Table 4) and the HAP emissions are summarized graphically (Figures 4 and 5) here. All emission data is presented in detail in electronic form in Appendix 2.

A summary of the kiln conditions for each sampling interval is shown in Table 4. A collection interval is the time the impingers were on and sampling occurred, approximately 90 minutes. An adjusted interval is the period spanning the midpoints between collection intervals, about six hours. For example, if the impingers were on from 12:00 to 13:30, 18:00 to 19:30, and 00:00 to 1:30, the 18:00 to 19:30 impinger set represents the adjusted interval from 15:45 to 21:45. The mass calculations are adjusted proportionally to represent emissions during the adjusted interval. For example, if a collection interval was 90 minutes and the adjusted interval was six hours, the amount of HAPs in the impinger is multiplied by four. Sampling occurred for approximately 28% of the drying time.

The MACT HAP emissions and the emissions of ethanol and acetic acid are shown in Table 5. The total HAP emissions were 0.13 lb/mbf for the red alder (does not include the non-HAPs, ethanol and acetic acid). Acetaldehyde and methanol were

emitted in nearly equal quantities, 0.062 and 0.063 lb/mbf, respectively. Other HAPs (formaldehyde, propionaldehyde, and acrolein) are present and comprise only 1.6% of all the HAPs. Phenol was not detected in any sample.

The HAP emissions as a function of time and wood moisture content during the cycle are shown in Figures 4 and 5, respectively. The rate of HAP emissions decreases with time throughout the schedule (lines are mostly concave downward in Figure 4). The rate of HAP emissions per percent moisture content change are greatest (lines in Figure 5 are concave upward) at low moisture content (even though they are lowest per unit of time because of the slow drying rate at low moisture content).

Sample	Interval	Interval	mass	Dry gas	Humidity	Con	tent
Run ID				flow rate		Mid	End
	hours	hours	kg	kg/min	mol/mol	%	%
1	1.50	3.66	10.358	0.047	0.057	106.1	105.5
2	1.50	5.96	62.682	0.175	0.126	102.7	99.2
3	1.52	5.96	18.363	0.051	0.268	98.1	95.1
4	1.57	6.06	31.383	0.086	0.287	91.9	87.7
5	1.83	6.06	80.956	0.223	0.253	79.2	70.9
6	1.52	5.96	81.109	0.227	0.243	62.8	54.6
7	1.48	5.96	68.084	0.190	0.242	47.9	41.0
8	1.55	6.01	52.652	0.146	0.242	35.1	30.5
9	1.55	6.01	31.904	0.089	0.242	26.9	24.1
10	1.55	6.01	28.697	0.080	0.241	21.2	18.5
11	1.52	5.96	17.525	0.049	0.240	16.5	15.0
12	1.52	6.01	10.285	0.029	0.240	14.0	12.9
13	1.57	6.01	8.801	0.024	0.239	11.9	11.2
14	1.50	6.01	6.384	0.018	0.239	10.5	9.9
15	1.48	6.01	7.921	0.022	0.238	9.5	8.4
16	1.50	1.85	2.224	0.020	0.236	7.8	8.0
SUM	1	89.47					

Table 4. Summary of HAP sampling intervals.

	Interval	Wood				Unit mass	leaving kiln			
Sample	Endpoint	Moisture	Methanol	Phenol	Ethanol	Acetic	Form-	Acet-	Propion-	Acrolein
Run ID		Content				acid	aldehyde	aldehyde	aldehyde	
	hours	%	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf
1	3.66	105.5	0.0000	0.0000	0.0004	0.0003	0.00001	0.0000	0.00000	0.00000
2	9.61	99.2	0.0008	0.0000	0.0315	0.0020	0.00006	0.0024	0.00002	0.00000
3	15.57	95.1	0.0026	0.0000	0.0836	0.0021	0.00003	0.0184	0.00004	0.00009
4	21.63	87.7	0.0057	0.0000	0.0786	0.0064	0.00004	0.0046	0.00003	0.00007
5	27.69	70.9	0.0050	0.0000	0.0553	0.0158	0.00008	0.0039	0.00003	0.00008
6	33.65	54.6	0.0084	0.0000	0.0630	0.0302	0.00010	0.0042	0.00004	0.00010
7	39.60	41.0	0.0065	0.0000	0.0435	0.0293	0.00011	0.0048	0.00006	0.00012
8	45.61	30.5	0.0067	0.0000	0.0304	0.0270	0.00006	0.0041	0.00004	0.00007
9	51.62	24.1	0.0050	0.0000	0.0232	0.0219	0.00005	0.0042	0.00004	0.00006
10	57.63	18.5	0.0054	0.0000	0.0127	0.0195	0.00004	0.0031	0.00003	0.00003
11	63.59	15.0	0.0048	0.0000	0.0070	0.0124	0.00005	0.0037	0.00004	0.00004
12	69.59	12.9	0.0020	0.0000	0.0042	0.0066	0.00002	0.0028	0.00003	0.00003
13	75.60	11.2	0.0014	0.0000	0.0037	0.0074	0.00002	0.0021	0.00003	0.00002
14	81.61	9.9	0.0031	0.0000	0.0024	0.0049	0.00002	0.0016	0.00002	0.00002
15	87.62	8.4	0.0044	0.0000	0.0028	0.0064	0.00005	0.0024	0.00008	0.00003
16	89.47	8.0	0.0003	0.0000	0.0001	0.0004	0.00000	0.0002	0.00001	0.00000
		Sums:	0.062	0.000	0.443	0.192	0.0007	0.063	0.0005	0.0007

Table 5. Summary of the HAP, acetic acid, and ethanol emissions.



Figure 4. HAP emissions as a function of time.



Figure 5. HAP emissions as a function of moisture content.

The detection limits for the GC instrument were

Methanol – 1.54  $\mu$ g/mL in the aqueous phase Phenol – 0.89  $\mu$ g/mL in the aqueous phase Ethanol – 0.98  $\mu$ g/mL in the aqueous phase Acetic acid – 4.85  $\mu$ g/mL in the aqueous phase Formaldehyde - 0.05  $\mu$ g/mL in the hexane phase Acetaldehyde – 0.05  $\mu$ g/mL in the hexane phase Propionaldehyde – 0.05  $\mu$ g/mL in the hexane phase Acrolein – 0.06  $\mu$ g/mL in the hexane phase

The method detection limit varies with gas flow through the impingers and the amount of solution in the impingers. Based on the flow conditions and impinger volumes for the 16 impinger samples, the method detection limits in the sampled gas (wet kiln exhaust) were

Methanol - 0.9-1.2 ppm Phenol - 0.2-0.34 ppm Ethanol – 0.4-0.56 ppm Acetic acid – 1.5-2.0 ppm

Formaldehyde – 0.01-0.02 ppm Acetaldehyde - 0.009 – 0.014 ppm Propionaldehyde - 0.006-0.01 ppm Acrolein - mean = 0.008-0.013 ppm

Methanol and acetic acid were below the detection limits in the first two samples. Acrolien was not detected in the first two samples, but was above detection limits in all the other samples. All other samples were above the detection limits. The table below shows the amounts emitted if one-half the detection limit is substituted for all samples below the detection limit. The total HAPs remain essentially unchanged at 0.127 lb/mbf.

			Unit mass	leaving kiln			
Methanol	Phenol	Ethanol	Acetic	Form-	Acet-	Propion-	Aarolain
Wethanor	Phenoi	Ethanol	acid	aldehyde	aldehyde	aldehyde	Acrolein
lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf
0.063	0.000	0.443	0.192	0.0007	0.063	0.0005	0.0007

Field spikes were run by operating two impinger trains simultaneously. An aliquot of the compounds was added to one impinger train. Spike recovery percentage is the mass of a compound detected in the lab compared to mass added to the impinger. Table 6 shows the field spike recoveries. The method requires between 70% and 130% recovery if the concentration of the compound in the gas phase is greater than 1.5 ppm in the dry gas. All spike recoveries were within +/- 30%. The spike levels (the amount in the spiked impinger should be between three to five times that in the unspiked impinger) were correct in at least one spike for methanol, ethanol, acetic acid and acetaldehyde. Formaldehyde, propionaldehyde, and acrolien were closest in the run 6 spike. We were attempting the "bracket method" with a high and low spike; however the concentrations in the catch were lower than anticipated. Recoveries were excellent, however.

The results for a field blank are shown in Table 7. None of the target compounds were detected in the blank above the detection limits. Formaldehyde was detected at approximately 10% of the detection limit and 20x lower than the typical sample concentration. Acetic acid was detected at approximately half of the detection limit and 10x lower than the typical sample concentration.

Duplicate samples were run by operating two impinger trains simultaneously. The results of duplicates are shown in Table 8. The percentage is the difference between the gas concentrations detected by each impinger. Phenol was not detected so duplicates could not be compared. Differences ranged from 2 to 26%, all within the limits of the method.

# Table 6. Spike test results.

		Mass in	n impinger	Alcohol Sp	Impinger	N N	Aass corre	ected for fi	ow
Run	Methano			Acetic	flow	Methano			Acetic
	þg	μg	hà	рд	mL/min	hà	рų	рд	64
6	205.2	0.0	1534.6	734.3	409.1	223.1	0.0	1668.0	798.2
602	507.1	61.2	4038.3	3406.3	444.6	507.1	61.2	4038.3	3406.3
			1						
Spike			ncentration		A. Barrelo			ecoveries	
mass	Methano		Ethanol	Acetic	States -	Methano		Ethanol	Acetic
9	µg/mL	µg/mL	µg/mL	µg/mL	C. C. C.	%	%	%	%
0.5	600.0	118.0	4753.2	5051.8	A State	94.7	103.6	99.7	103.3
			1	1	1	1 01.1	1.00.0	1 00.1	1 100.0
									***************
			Aldehyde S	ipike					a
			impinger		Impinger			ected for fl	w
Run	Form-	Acet-	Propion-		flow	Form-	Acet-	Propion-	Acroleir
	aldehyde		aldehyde				aldehyde	aldehyde	
	hà	hà	hà	рд	mL/min	hð	hà	Рð	Pha Pa
6	2.5	103.4	1.0	2.4	409.1	1.0	41.3	0.4	1.0
603	169.2	772.0	28.6	102.8	163.5	169.2	772.0	28.6	102.8
	1	Spike con	centrations		The second second	-	Coiko r		
Spike mass	Form-	Acet-	Propion	, 	A MARCE	Form		ecoveries	
abure mass	aldehyde			Acrolein	<b>的</b> 新的		Acet-	Propion-	Acroleir
6					100000	aldehyde			
<u>9</u> 0.57	µg/mL 369.5	µg/mL 1014.0	µg/mL	µg/mL	Contraction of the	%	%	%	%
0.01	009.0	1014.0	91.5	159.3	NR. * 108 **10*	79.8	126.4	54.0	112.2
			1						*****************
			-	Alcohol S-	ike				
	1	Mass in	impinger	Alcohol Sp	Impinger	L N	ass corre	cted for fk	
Run	Methanol	Phenol	Ethanol	Acetic	flow	Methanol		Ethanol	Acetic
		Phenor			mL/min				
9	<u>µg</u> 315,4	0.0	µg 1469.9	µg 1387.5	409.0	µg 144.6	<u>94</u> 0.0	674 O	Pg
903	802.2	134.6	5680.7	6140.7	187.5	802.2	134.6	674.0	636.2
	004.4	104.0	1 0000.7	0140.7	107.5	002.2	1.134.0	5680.7	6140.7
Spike	1	Spike con	centrations		ALL	r	Spike re	coveries	
mass	Methanol	Phenol	Ethanol	Acetic	and the	Methanol		Ethanol	Acetic
g	µg/mL	µg/mL	µg/mL	µg/mL		%	%	%	%
Sector D.V.	1. S. S. S. S.	STATES	100000-0		1999	1. Second	The SPECIES		and the second
1.12	600.0	118.0	4753.2	5051.8	A GUNA	97.9	101.8	94.0	97.3
	1		Idebude C	oiko					
			Idehyde S						
		Mass in		pinte		M	ass corre	cled for fly	w
D.	Form-	Mass in Acet-	impinger		Impinger			cted for flo	
Run		Acet-	Impinger Propion-	Acrolein	Impinger flow	Form-	Acet-	Propion-	
Run	aldehyde	Acet- aldehyde	Impinger Propion- aldehyde	Acrolein	flow	Form- aldehyde	Acet- aldehyde	Propion- aldehyde	Acrolein
Run 9	aldehyde µg	Acet- aldehyde µg	Impinger Propion- aldehyde µ9	Acrolein µg	flow mL/min	Form- aldehyde µg	Acet- aldehyde µg	Propion- aldehyde µg	Acrolein µg
	aldehyde	Acet- aldehyde	Impinger Propion- aldehyde	Acrolein	flow	Form- aldehyde µg 3.3	Acet- aldehyde µg 291.1	Propion- aldehyde µg 2.6	Acrolein µg 4.0
9	aldehyde µg 3.0	Acet- aldehyde µg 265.1	Impinger Propion- aldehyde Pg 2.3	Acrolein µg 3.7	flow mL/min 409.0	Form- aldehyde µg	Acet- aldehyde µg	Propion- aldehyde µg	Acrolein µg
9 902	aldehyde µg 3.0 328.6	Acet- aldehyde µg 265.1 1209.4	Impinger Propion- aldehyde Pg 2.3	Acrolein µg 3.7 158.7	flow mL/min 409.0	Form- aldehyde µg 3.3	Acet- aldehyde µg 291.1 1209.4	Propion- aldehyde µg 2.6	Acrolein µg 4.0
9 902	aldehyde µg 3.0 328.6	Acet- aldehyde µg 265.1 1209.4	impinger Propion- aldehyde ¥9 2.3 87.0	Acrolein µg 3.7 158.7	flow mL/min 409.0	Form- aldehyde µg 3.3	Acet- aldehyde µg 291.1 1209.4	Propion- aldehyde µg 2.6 87.0	Acrolein µg 4.0 158.7
9 902	aldehyde µg 3.0 328.6	Acet- aldehyde µg 265.1 1209.4 Spike con	impinger Propion- aldehyde ¥9 2.3 87.0 centrations	Acrolein µg 3.7 158.7	flow mL/min 409.0	Form- aldehyde ¥9 3.3 328.6	Acet- aldehyde µg 291.1 1209.4 Spike re Acet-	Propion- aldehyde µg 2.6 87.0 coveries	Acrolein µg 4.0
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9 902 Spike mass	aldehyde µg 3.0 328.6 Form- aldehyde	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde	impinger Propion- aldehyde µg 2.3 87.0 centrations Propion- aldehyde	Acrolein µg 3.7 158.7 Acrolein	flow mL/min 409.0	Form- aldehyde µg 3.3 328.6 Form- aldehyde	Acet- aldehyde µg 291.1 1209.4 Spike re Acet- aldehyde	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde	Acrolein µg 4.0 158.7 Acrolein
9 902 Spike mass 9	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL	impinger Propion- aldehyde µg 2.3 87.0 centrations Propion- aldehyde µg/mL	Acrolein µg 3.7 158.7 Acrolein µg/mL	flow mL/min 409.0	Form- aldehyde µg 3.3 328.6 Form- aldehyde %	Acet- aldehyde µg 291.1 1209.4 Spike re Acet- aldehyde %	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde %	Acrolein µg 4.0 158.7 Acrolein %
9 902 Spike mass 9	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL	Impinger Propion- aldehyde <u>Pg</u> 2.3 87.0 centrations Propion- aldehyde <u>µg/mL</u> 91.5	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3	flow mL/min 409.0 449.1	Form- aldehyde µg 3.3 328.6 Form- aldehyde %	Acet- aldehyde µg 291.1 1209.4 Spike re Acet- aldehyde %	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde %	Acrolein µg 4.0 158.7 Acrolein %
9 902 Spike mass 9	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0	impinger Propion- aldehyde µg 2.3 87.0 centrations Propion- aldehyde µg/mL 91.5	Acrolein µg 3.7 158.7 Acrolein µg/mL	flow mL/min 409.0 449.1	Form- aldehyde µg 3.3 328.6 Form- aldehyde % 82.3	Acet- aldehyde µg 291.1 1209.4 Spike re Acet- aldehyde % 84.6	Propion- aldehyde µg 2.6 87,0 ecoveries Propion- aldehyde % 86,2	Acrolein µg 4.0 158.7 Acrolein % 90.7
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9 902 Spike mass <u>9</u> 1.07 Run 13	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0	Impinger Propion- aldehyde <u>µg</u> 2.3 87.0 centrations Propion- aldehyde <u>µg/mL</u> 91.5 <i>Palse</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i> <i>pg</i>	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 Vcohol Spi Acetic <u>µg</u> 1712.2	flow mL/min 409.0 449.1 449.1 ke Impinger flow mL/min 410.7	Form- aldehyde µg 33 328.6 Form- aldehyde % 82.3 Methanol µg 367.3	Acet- aldehyde µg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 Phenol µg 0.0	Propion- aldehyde µg 2.6 87.0 ecoveries Propion- aldehyde % 86.2 etced for flo Ethanol µg 951.9	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 w Acetic <u>µg</u> 1882.4
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9 902 Spike mass 9 1.07 Run 13 1303	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 1681.7	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7	impinger Propion- aldehyde <u>µg</u> 2.3 87.0 centrations Propion- aldehyde <u>µg/mL</u> 91.5 <i>Propion- aldehyde</i> <u>µg/mL</u> 91.5 <i>Propion- aldehyde</i> <u>µg/mL</u> 91.5 <i>Propion- aldehyde</i> <u>µg/mL</u> 91.5 <i>Propion- aldehyde</i> <u>µg/mL</u> 91.5 <i>Propion- aldehyde</i> <u>µg/mL</u> 91.5 <i>Propion- aldehyde</i> <u>µg/mL</u> 91.5 <i>Propion- aldehyde</i> <u>µg/mL</u> 91.5 <i>Propion- aldehyde</i> <u>µg/mL</u> 91.5 <i>Propion- aldehyde</i> <i>µg/mL</i> <i>Propion- aldehyde</i> <i>µg/mL</i> <i>Propion- aldehyde</i> <i>µg/mL</i> <i>Propion- aldehyde</i> <i>µg/mL</i> <i>Propion- aldehyde</i> <i>µg/mL</i> <i>Propion- <i>propion- aldehyde</i> <i>µg/mL</i> <i>Propion- <i>propion- aldehyde</i> <i>Propion- <i>propion- aldehyde</i> <i>Propion- <i>propion- aldehyde</i> <i>Propion- <i>propion- aldehyde</i> <i>Propion- <i>propion- <i>propion- <i>propion- <i>propion- <i>propion- <i>propion- <i>propion- <i>propion- <i>propion- <i>propion- <i>propion-</i> <i>propion- <i>propion-</i> <i>propion- <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propion-</i> <i>propio</i></i></i></i></i></i></i></i></i></i></i></i></i></i></i></i></i></i></i>	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 Vcohol Spi Acetic <u>µg</u> 1712.2	flow mL/min 409.0 449.1 449.1 ke Impinger flow mL/min 410.7	Form- aldehyde µg 33 328.6 Form- aldehyde % 82.3 Methanol µg 367.3	Acet- aldehyde µg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 ass corre Phenol µg 0.0 243.7	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol µg 951.9 10103.7	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 w Acetic <u>µg</u> 1882.4
9 902 Spike mass <u>9</u> 1.07 Run 13 1303 Spike	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 1681.7	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike con	impinger Propion- aldehyde µg 2.3 87.0 centrations Propion- aldehyde µg/mL 91.5	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 Nochol Spi Acetic <u>µg</u> 1712.2 11566.8	flow mL/min 409.0 449.1 449.1 ke Impinger flow mL/min 410.7	Form- aldehyde µg 3.3 328.6 Form- aldehyde % 82.3 Methanol µg 367.3 1681.7	Acet- aldehyde µg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 84.6 Sass corre- Phenol µg 0.0 243.7 Spike re	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 86.2 cted for flo Ethanol µg 951.9 10103.7 coveries	Acrolein 4.0 158.7 Acrolein % 90.7 w Acetic µg 1882.4 11566.8
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9 902 Spike mass 9 1.07 Run 13 1303 Spike	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 1681.7	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike con	impinger Propion- aldehyde µg 2.3 87.0 centrations Propion- aldehyde µg/mL 91.5	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 Nochol Spi Acetic <u>µg</u> 1712.2 11566.8	flow mL/min 409.0 449.1 449.1 ke Impinger flow mL/min 410.7	Form- aldehyde µg 3.3 328.6 Form- aldehyde % 82.3 Methanol µg 367.3 1681.7	Acet- aldehyde µg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 84.6 Sass corre- Phenol µg 0.0 243.7 Spike re	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 86.2 cted for flo Ethanol µg 951.9 10103.7 coveries	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 w Acetic <u>µg</u> 1882.4 11566.8
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 1681.7 Methanol	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL	impinger Propion- aldehyde µg 2.3 87.0 centrations Propion- aldehyde µg/mL 91.5	Acrolein Jg 3.7 158.7 Acrolein Jg/mL 159.3 Acrolein Jg/mL 159.3 Acetic Jg 1712.2 11566.8 Acetic Jg/mL	flow mL/min 409.0 449.1 449.1 ke Impinger flow mL/min 410.7	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol yg 367.3 1681.7 Methanol %	Acet- aldehyde yg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 Acet- aldehyde % 84.6 Phenol yg 0.0 0.2 43.7 Spike re Phenol %	Propion- aldehyde J9 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol J9 951.9 10103.7 coveries Ethanol %	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 <u>90.7</u> <u>w</u> <u>Acetic</u> <u>µg</u> 1882.4 11566.8 <u>Acetic</u> %
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg Methanol µg/mL	Acet- aldehyde µg 265.1 1209.4 Spike con- Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike con-	impinger Propion- aldehyde J9 87.0 centrations Propion- aldehyde µg/mL 91.5 91.5 Propion- aldehyde µg/mL Ethanol	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 Voohol Spi Acetic <u>µg</u> 1712.2 11566.8 Acetic	flow mL/min 409.0 449.1 449.1 ke Impinger flow mL/min 410.7	Form- aldehyde µg 3.3 328.6 Form- aldehyde % 82.3 Methanol µg 367.3 1681.7 Methanol	Acet- aldehyde µg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 Phenol µg 0.0 243.7 Spike re Phenol	Propion- aldehyde yg 2.6 87.0 ecoveries Propion- aldehyde % 86.2 ecoveries thanol yg 951.9 10103.7 ecoveries Ethanol	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 w Acetic <u>µg</u> 1882.4 11566.8 Acetic
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg Methanol µg/mL	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL 118.0	impinger Propion- aldehyde <u>µg</u> 2.3 87.0 87.0 87.0 91.5 91.5 91.5 <i>Propion- aldehyde µg/mL</i> 91.5 91.5 91.5 91.5 91.5 91.5 91.5 91.5	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 <u>Vcohol Spi</u> Acetic <u>µg</u> 1712.2 11566.8 Acetic <u>µg/mL</u> 5051.8	flow mL/min 409.0 449.1 449.1 ke Impinger flow mL/min 410.7	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol yg 367.3 1681.7 Methanol %	Acet- aldehyde yg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 Acet- aldehyde % 84.6 Phenol yg 0.0 0.2 43.7 Spike re Phenol %	Propion- aldehyde J9 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol J9 951.9 10103.7 coveries Ethanol %	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 90.7 w Acetic <u>µg</u> 1882.4 11566.8 Acetic %
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg Methanol µg/mL	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL 118.0	impinger Propion- aldehyde J9 87.0 centrations Propion- aldehyde µg/mL 91.5 Propion- aldehyde µg/mL 91.5 Propion- aldehyde µg/mL 91.5 Propion- aldehyde µg/mL 4753.2 Idehyde Sp	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 <u>Vcohol Spi</u> Acetic <u>µg</u> 1712.2 11566.8 Acetic <u>µg/mL</u> 5051.8	flow mL/min 409.0 449.1 449.1 ke Impinger flow mL/min 410.7	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol yg 367.3 1681.7 Methanol % 106.9	Acet- aldehyde yg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 Acet- aldehyde % 84.6 Phenol yg 0,0 243.7 Spike re Phenol y0,0 243.7	Propion- aldehyde yg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol yg 951,9 10103.7 coveries Ethanol % 93.9	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 ww Acetic <u>µg</u> 1882.4 11566.8 Acetic % 93.5
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 1681.7 Methanol µg/mL 600.0	Acet- aldehyde µg 265.1 1209.4 Spike con- Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL 118.0 A Mass in	impinger Propion- aldehyde yg 2.3 87.0 centrations Propion- aldehyde µg/mL 91.5 91.5 91.5 91.5 91.5 91.5 91.5 91.5	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 <u>Vcohol Spi</u> Acetic <u>µg</u> 1712.2 11566.8 Acetic <u>µg/mL</u> 5051.8	flow <u>mUrnin</u> 409.0 449.1 449.1 ke Impinger flow mU/min 410.7 451.6	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol yg 367.3 1681.7 Methanol % 106.9	Acet- aldehyde yg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 Acet- aldehyde % 84.6 Phenol yg 0.0 243.7 Spike re Phenol % 100.7	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for filo Ethanol µg 951.9 10103.7 coveries Ethanol % 93.9	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 ww Acetic <u>µg</u> 1882.4 11566.8 Acetic % 93.5
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9 2.05	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg/mL 600.0 Form-	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL 118.0 Assin Acet-	impinger Propion- aldehyde µg 2.3. 87.0 centrations Propion- aldehyde µg/mL 91.5 <i>Propion- aldehyde</i> µg/mL 91.5 <i>Propion- aldehyde</i> µg/mL 4753.2 Idehyde Sp impinger Propion-	Acrolein <u>J9</u> 3.7 158.7 Acrolein <u>J99/mL</u> 159.3 Jcohol Spi Acetic <u>J99/mL</u> 5051.8 Sike	flow mL/min 409.0 449.1 449.1 ke Impinger flow mL/min 410.7 451.6	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol yg 367.3 1681.7 Methanol % 106.9	Acet- aldehyde yg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 ass correc Phenol yg 0.0 0.243.7 Spike re Phenol yg 100.7	Propion- aldehyde yg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol yg 951.9 10103.7 coveries Ethanol % 93.9	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 ww Acetic <u>µg</u> 1882.4 11566.8 Acetic % 93.5 ww
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 Methanol µg/mL 600.0	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike con Phenol µg/mL 118.0 A Mass in Acet- aldehyde	impinger Propion- aldehyde yg 2.3 87.0 centrations Propion- aldehyde µg/mL 91.5 91.5 91.5 91.5 91.5 91.5 91.5 91.5	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 <u>Vcohol Spi</u> Acetic <u>µg</u> 1712.2 11566.8 Acetic <u>µg/mL</u> 5051.8	flow mUmin 409.0 449.1 449.1 ke Impinger flow Impinger flow	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol yg 367.3 1681.7 Methanol % 106.9	Acet- aldehyde yg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 ass correc Phenol yg 0.0 0.243.7 Spike re Phenol yg 100.7	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for filo Ethanol µg 951.9 10103.7 coveries Ethanol % 93.9	Acrolein <u>µg</u> 4.0 158.7 Acrolein % 90.7 ww Acetic <u>µg</u> 1882.4 11566.8 Acetic % 93.5
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9 2.05 Run	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 1681.7 Methanol µg/mL 600.0	Acet- aldehyde µg 265.1 1209.4 Spike con- Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike con- Phenol µg/mL 118.0 Acet- aldehyde µg/g	impinger Propion- aldehyde yg 2.3 87.0 centrations Propion- aldehyde yg/mL 91.5 91.5 91.5 91.5 91.5 91.5 91.5 91.5	Астоlein <u>µg</u> 3.7 158.7 Астоlein <u>µg/mL</u> 159.3 Nocohol Spi Acetic <u>µg</u> 1712.2 11566.8 Асеtic <u>µg/mL</u> 5051.8 storthere Solorin <u>µg/mL</u> 159.3 Nocohol Spi 159.3 Nocohol Spi 159.5 Nocohol Spi 159.5 Nocoho	flow mU/min 409.0 449.1 449.1 ke Impinger flow mL/min flow mL/min	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol % 367.3 1681.7 Methanol % 106.9 M. Form- aldehyde yg	Acet- aldehyde yg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 Acet- aldehyde yg 100.7 Acet- aldehyde yg	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol µg 951.9 10103.7 coveries Ethanol % 93.9 coveries	Acrolein Hg 4.0 158.7 Acrolein % 90.7 W Acrolein Hg 1882.4 11566.8 Acetic % 93.5 W Acrolein Hg Hg Hg Hg
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9 2.05 Run 13	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 1681.7 Methanol µg/mL 600.0 Form- aldehyde µg/s.3 Methanol µg/mL 600.0	Acet- aldehyde µg 265.1 1209.4 Spike con- Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL 118.0 Acet- aldehyde µg/mL 118.0	impinger Propion- aldehyde µg 2.3 87.0 centrations Propion- aldehyde µg/mL 91.5 <i>Propion- aldehyde</i> µg/mL 10103.7 totnar.2 4753.2 Idehyde Sp impinger Propion- aldehyde µg 9.6.6	Acrolein <u>µg</u> 158.7 Acrolein <u>µg/mL</u> 159.3 Jochol Spi Acetic <u>µg</u> 1712.2 11566.8 Acetic <u>µg/mL</u> 5051.8 Jike Acrolein <u>µg/mL</u> 4.3	flow mUmin 409.0 449.1 449.1 ke Impinger flow mUmin 410.7 451.6	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol yg 367.3 1681.7 Methanol % 106.9 No.9	Acet- aldehyde yg 291,1 1209,4 Spike re Acet- aldehyde % 84.6 % 84.6 % Phenol yg 0.0 0.243.7 Spike re Phenol % 100.7 %	Propion- aldehyde yg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol yg 951.9 10103.7 coveries Ethanol % 93.9 coveries Stanol % 93.9 coveries Stanol St	Acrolein <u>yg</u> 4.0 158.7 Acrolein <u>%</u> 90.7 <u>ww</u> Acetic <u>yg</u> 1882.4 Acetic <u>%</u> 93.5 <u>ww</u> Acrolein <u>yg</u> 2.0
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9 2.05 Run	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 1681.7 Methanol µg/mL 600.0	Acet- aldehyde µg 265.1 1209.4 Spike con- Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike con- Phenol µg/mL 118.0 Acet- aldehyde µg/g	impinger Propion- aldehyde yg 2.3 87.0 centrations Propion- aldehyde yg/mL 91.5 91.5 91.5 91.5 91.5 91.5 91.5 91.5	Астоlein <u>µg</u> 3.7 158.7 Астоlein <u>µg/mL</u> 159.3 Nocohol Spi Acetic <u>µg</u> 1712.2 11566.8 Асеtic <u>µg/mL</u> 5051.8 storthere Solorin <u>µg/mL</u> 159.3 Nocohol Spi 159.3 Nocohol Spi 159.5 Nocohol Spi 159.5 Nocoho	flow mU/min 409.0 449.1 449.1 ke Impinger flow mL/min flow mL/min	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol % 367.3 1681.7 Methanol % 106.9 M. Form- aldehyde yg	Acet- aldehyde yg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 Acet- aldehyde yg 100.7 Acet- aldehyde yg	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol µg 951.9 10103.7 coveries Ethanol % 93.9 coveries	Acrolein Hg 4.0 158.7 Acrolein % 90.7 W Acrolein Hg 1882.4 11566.8 Acetic % 93.5 W Acrolein Hg Hg Hg Hg
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9 2.05 Run 13	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg/mL 600.0 Form- aldehyde µg/mL 603.0 S.3 403.5	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL 118.0 A Mass in Acet- aldehyde µg/mL 118.0	impinger Propion- aldehyde µg 2.3 87.0 S7.0 S7.0 S7.0 S7.0 S7.0 Propion- aldehyde µg/mL 91.5 S S S S S S S S S S S S S S S S S S S	Acrolein <u>µg</u> 158.7 Acrolein <u>µg/mL</u> 159.3 Jochol Spi Acetic <u>µg</u> 1712.2 11566.8 Acetic <u>µg/mL</u> 5051.8 Jike Acrolein <u>µg/mL</u> 4.3	flow mUmin 409.0 449.1 449.1 ke Impinger flow mUmin 410.7 451.6	Form- aldehyde yg 3.3 328.6 Form- aldehyde % 82.3 Methanol yg 367.3 1681.7 Methanol % 106.9 No.9	Acet- aldehyde yg 291.1 1209.4 Spike re Acet- aldehyde % 84.6 ass corree Phenol yg 0.0 243.7 Spike re Phenol yg 100.7 spike re Acet- aldehyde yg 100.7	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol µg 951.9 10103.7 coveries Ethanol % 93.9 sted for flo Propion- aldehyde µg 3.1 85.9	Acrolein <u>yg</u> 4.0 158.7 Acrolein <u>%</u> 90.7 <u>ww</u> Acetic <u>yg</u> 1882.4 Acetic <u>%</u> 93.5 <u>ww</u> Acrolein <u>yg</u> 2.0
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9 2.05 Run 13 1302	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 1681.7 Methanol µg/mL 600.0 Form- aldehyde µg 5.3 403.5	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL 118.0 Acet- aldehyde µg/mL 118.0 Acet- aldehyde µg/mL 118.0 Acet- aldehyde µg/mL 118.0 Acet- aldehyde µg/mL 118.0 Acet- aldehyde µg/mL 118.0 Acet- aldehyde µg/mL 118.0 Acet- aldehyde µg/mL	impinger Propion- aldehyde µg 2.3 87.0 centrations Propion- aldehyde µg/mL 91.5 £thanol µg/mL 4753.2 Idehyde Sp impinger Propion- aldehyde g/mL 4753.2 Idehyde Sp impinger Propion- aldehyde §6.6 85.9 centrations	Acrolein <u>µg</u> 158.7 Acrolein <u>µg/mL</u> 159.3 Jochol Spi Acetic <u>µg</u> 1712.2 11566.8 Acetic <u>µg/mL</u> 5051.8 Jike Acrolein <u>µg/mL</u> 4.3	flow mUmin 409.0 449.1 449.1 ke Impinger flow mUmin 410.7 451.6	Form- aldehyde µg 3.3 328.6 Form- aldehyde % 82.3 Methanol µg 367.3 1681.7 Methanol % 106.9 106.9	Acet- aldehyde yg 291,1 1209,4 Spike re Acet- aldehyde % 84.6 ass correc Phenol yg 0.0 243.7 Spike re Phenol % 100.7 ass correc Acet- aldehyde yg 226.5 2189.3	Propion- aldehyde Jg 2.6 87.0 coveries Propion- aldehyde % 86.2 cted for flo Ethanol Jg 951.9 10103.7 coveries Ethanol % 93.9 coveries cted for flo Propion- aldehyde Jg 93.9 coveries coveries cted for flo Propion- aldehyde Jg 93.9 coveries covere	Acrolein <u>yg</u> 4.0 158.7 Acrolein <u>%</u> 90.7 <u>ww</u> Acetic <u>yg</u> 1882.4 Acetic <u>%</u> 93.5 <u>ww</u> Acrolein <u>yg</u> 2.0
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9 2.05 Run 13 1302	aldehyde <u>µg</u> 3.0 328.6 Form- aldehyde <u>µg/mL</u> 369.5 Methanol <u>µg</u> 334.1 1681.7 Methanol <u>µg/mL</u> 600.0	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL 118.0 Acet- aldehyde µg/at.2 2189.3 Spike conc Acet-	impinger Propion- aldehyde J9 87.0 centrations Propion- aldehyde µg/mL 91.5 91.5 91.5 91.5 91.5 91.5 91.5 91.5	Acrolein <u>µg</u> 158.7 Acrolein <u>µg/mL</u> 159.3 Jochol Spi Acetic <u>µg</u> 1712.2 11566.8 Acetic <u>µg/mL</u> 5051.8 Jike Acrolein <u>µg/mL</u> 4.3	flow mU/min 409.0 449.1 449.1 ke Impinger flow mU/min 410.7 451.6 Impinger flow mL/min 410.7 193.3	Form- aldehyde µg 3.3 328.6 Form- aldehyde % 82.3 M Methanol µg 367.3 1681.7 Methanol % 106.9 N. Form- aldehyde 2.5 403.5	Acet- aldehyde yg 291,1 1209,4 Spike re Acet- aldehyde % 84.6 ass correc Phenol % 0.0 243.7 Spike re Acet- aldehyde yg 226.5 2189.3 Spike re Acet-	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 coveries Ethanol % 93.9 coveries coveries coveries coveries propion- aldehyde µg 3.1 85.9 coveries	Acrolein <u>yg</u> 4.0 158.7 Acrolein <u>%</u> 90.7 w Acrolein <u>1882.4</u> 11566.8 <u>Acetic</u> <u>%</u> 93.5 w Acrolein <u>yg</u> 2.0 280.4
9 902 Spike mass 9 1.07 Run 13 1303 Spike mass 9 2.05 Run 13	aldehyde µg 3.0 328.6 Form- aldehyde µg/mL 369.5 Methanol µg 334.1 1681.7 Methanol µg/mL 600.0 Form- aldehyde µg 5.3 403.5	Acet- aldehyde µg 265.1 1209.4 Spike con Acet- aldehyde µg/mL 1014.0 Mass in Phenol µg 0.0 243.7 Spike conc Phenol µg/mL 118.0 Acet- aldehyde µg/at.2 2189.3 Spike conc Acet-	impinger Propion- aldehyde µg 2.3 87.0 centrations Propion- aldehyde µg/mL 91.5 £thanol µg/mL 4753.2 Idehyde Sp impinger Propion- aldehyde yg/mL 4753.2 Idehyde Sp impinger Propion- aldehyde §6.6 85.9 centrations	Acrolein <u>µg</u> 3.7 158.7 Acrolein <u>µg/mL</u> 159.3 Nocohol Spi Acetic <u>µg</u> 1712.2 11566.8 Acetic <u>µg/mL</u> 5051.8 So51.8 Acrolein <u>µg</u> 4.3 280.4	flow mU/min 409.0 449.1 449.1 ke Impinger flow mU/min 410.7 451.6 Impinger flow mL/min 410.7 193.3	Form- aldehyde µg 3.3 328.6 Form- aldehyde % 82.3 Methanol µg 367.3 1681.7 Methanol % 106.9 106.9	Acet- aldehyde yg 291,1 1209,4 Spike re Acet- aldehyde % 84.6 ass correc Phenol % 0.0 243.7 Spike re Acet- aldehyde yg 226.5 2189.3 Spike re Acet-	Propion- aldehyde µg 2.6 87.0 coveries Propion- aldehyde % 86.2 coveries Ethanol % 93.9 coveries coveries coveries coveries propion- aldehyde µg 3.1 85.9 coveries	Acrolein <u>yg</u> 4.0 158.7 Acrolein <u>%</u> 90.7 <u>ww</u> Acetic <u>yg</u> 1882.4 Acetic <u>%</u> 93.5 <u>ww</u> Acrolein <u>yg</u> 2.0

Cascade Hardwoods, Red alder

		Field	d blank				FB
Methanol	Phenol	Ethanol	Acetic	Form-	Acet-	Propion-	Aaralain
Methanoi	Phenor	Ethanoi	acid	aldehyde	aldehyde	aldehyde	Acrolein
ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm
0.000	0.000	0.000	1.590	0.006	0.000	0.000	0.000

Table 7.	Results t	for the field	d blank.

 Table 8.
 Results for duplicate runs.

				Duplicate					
				Mass in	impinger				Impingo
Run	Methanol	Phenol	Ethanol	Acetic acid	Form- aldehyde	Acet- aldehyde	Propion- aldehyde	Acrolein	Impinge flow
	μg	μg	μg	μg	μg	μg	μg	μg	mL/min
5	144.8	0.0	1611	459.1	2.2	112.4	0.9	2.3	404.3
502	154.5	0.0	1629	461.3	2.4	129.0	1.1	2.6	444.0
Difference, %	2.9	#DIV/0!	8.3	8.9	2.2	4.4	9.1	5.2	1
				Duplicate					
				Mass in	impinger				Impipgo
Run	Methanol	Phenol	Ethanol	Acetic acid	Form- aldehyde	Acet- aldehyde	Propion- aldehyde	Acrolein	Impinge flow
	μg	μg	μg	μg	μg	μg	μg	μg	mL/min
10	376.5	0.0	888.5	1364.4	2.9	218.8	2.2	2.1	407.7
1002	494.8	0.0	1033.5	1633.2	3.0	240.9	2.4	4.5	444.7
Difference, %	18.6	#DIV/0!	6.4	9.3	6.0	0.9	4.1	65.3	
				Duplicate	Ð				-
- 1a3				Mass in	impinger				Impingo
Run	Methanol	Phenol	Ethanol	Acetic acid	Form- aldehyde	Acet- aldehyde	Propion- aldehyde	Acrolein	Impinge flow
	μg	μg	μg	μg	μg	μg	μg	μg	mL/min
14	968.8	0.0	725.9	1512.5	6.4	482.1	7.4	4.7	412.6
1402	1329.1	0.0	983.8	2128.1	6.2	537.7	8.3	5.2	447.8
Difference, %	23.3	#DIV/0!	22.1	25.8	12.7	2.7	3.8	0.8	

# 4. Control system and operating conditions

A schematic of the kiln is shown in Figure 6 (top). The kiln box is approximately 4' by 4' by 4'. It is indirectly heated by steam. Four dry-bulb thermocouples and two wet-bulb thermocouples are located on the entering-air side of the load. The dry-bulb thermocouples are spaced in a grid. The two wet-bulb thermocouples are under a single sock at the center of the entering-air side of the load.

## Humidity control

A 200 L/min MKS mass flow meter controlled the amount of air entering the kiln. It was factory calibrated and checked using a bubble meter. The amount of air entering the kiln is based on the wet-bulb temperature - if it is above setpoint, the airflow is increased and if it is below setpoint the airflow is decreased. This is analogous to venting for a commercial kiln. A minimum of 2 L/min entered the kiln at all times, more than removed by the analyzer (1.6 L/min). Putting air into the kiln at a rate of 100 L/min causes the pressure in the kiln to be 60 to 130 Pa above ambient, depending on location in the kiln (high-pressure or low-pressure side). Thus, any fugitive leakage should be out of the kiln. Two additional flow meters can be manually set to provide additional airflow.

## Temperature control

Temperature in the kiln is controlled by indirect steam heating. When the dry-bulb temperature is below setpoint, the steam pressure in the coil is increased. When it is above setpoint, steam flow to the coil is reduced.

The dry- and wet-bulb temperatures recorded for each charge are shown in Figure 7. The schedule provided by the mill is also shown.



**Figure 6.** Schematic of kiln and sampling system (top) and photo of charge in kiln (bottom).

# 5. Production-related parameters

#### Wood quantity

The wood properties were determined using the nominal wood dimensions (5/4 in this case) which provides for 1.25 board feet per square foot of board face. There were 42 pieces in the kiln at 44" in length. The sum of the 42 board widths was 226 inches. The board footage was therefore 90.04 board feet. This quantity was used to express the emissions from the drying cycle on a production basis of lb/mbf (pounds per thousand board feet).

#### Wood properties

The wood property measurements are shown in Table 9. Individual measurements can be found in the Excel file "Weights, Cascade.XLS" in Appendix 2.

Occasional tension wood was noted. The amount seemed typical for red alder.

Heartwood percentage was not determined because is cannot be easily distinguished from sapwood in red alder.

The average ring count was determined by counting the rings over a 1" radial distance and averaging for all boards.

The number of knots were counted on the top face of each board and averaged. Knot diameter is an average of the knots present. The knots occupied approximately 0.3% of the boards' faces.

Kn	ots	Heartwood	Ring	Pith
Number	Diameter		Count	In
#	in	%	#/inch	Y/N
2.3	0.6	N/D	7.9	3

Table 9. Wood properties.





# 6. Test methods

#### Charge Sequence

The lumber was unwrapped and 2" were trimmed from each end of each board to give 44" samples. These were then weighed, placed in the kiln and dried. At the end of drying the wood was weighed, oven dried, and reweighed so initial and final moisture contents could be determined by ASTM D4442 (oven-dry method).

# Sampling Methodologies

#### Hydrocarbon

Sampling for total hydrocarbon is done directly from the kiln as shown in Figure 6 (top). The concentration obtained from the hydrocarbon analyzer and the amount of air entering the kiln allow the total hydrocarbon emissions to be calculated.

Figure 8 shows the hydrocarbon sampling system. Unlike stack testing, all necessary equipment is located in a lab and flows are controlled with valves. The sample is withdrawn from the kiln under the assumption that the gas in the kiln is well-mixed and that the composition in the kiln near the exhaust is the same as the composition of the exhaust. The THC sample was drawn from the kiln into a heated dilution/filter box. The box was heated to 235-245°F. Heated dilution gas was added to the hydrocarbon sample gas to lower the gas moisture content to the detector (except for runs 1 and 2) so that the air moisture content to the detector remained less than 15%. The sample line from the box to the analyzer was heated to 245°F.

The fuel gas was hydrogen. The span gas was EPA Protocol 197 ppm propane in air, the mid-gas was EPA Protocol 50 ppm propane in air. The zero gas was <0.1 ppm air. Detailed sampling procedures are in Appendix 1.



**Figure 8.** Schematic of heated filter box with air dilution system, heated sample line, and analyzer (top). Sample enters heated box from back of drawing through a heated sampling line. Calibration gas valves and dilution air valves are to left. Line to analyzer is the green line on the left in the left photo.

# HAPs

The sampling train for NCASI Method 105 is shown in Figure 9. The impingers were in a glycol solution maintained at -1 C. Prior to each sampling interval, the impingers were laboratory-washed and 10 to 15 mL of BHA solution were added to the first and second impingers. The third impinger was left empty. The fourth impinger was present in the system to prevent any overflow from reaching the critical orifice. The system was then assembled and a vacuum check was performed with the valves at each end closed. Less than 1" Hg of pressure change over 2 minutes was acceptable. This was met for each interval. The flow rate through the system was then measured using a Gilibrator flow meter to take four flow readings at the probe tip. This was approximately 200-500 mL/min, depending on the sampling train. A valve at the probe tip was then turned to sample from the kiln and the sampling interval begun. The collection interval time was approximately 1:30 and an interval was started approximately every six hours.

The flow rate was measured after each sampling interval. The fluid in the three impingers was weighed and placed in a glass bottle. The impingers were then rinsed with 10 mL of water followed by 3 to 5 mL of hexane. The rinses were also placed in the bottle and it was sealed. Samples were kept refrigerated and in the dark until lab analysis was done. Lab analysis was done within one week of sample collection.

The local airport altimeter setting and the lab temperature were recorded at the beginning and end of each interval so the flow rates could be adjusted to standard conditions.



Figure 9. HAPs sampling train.

# 7. Analytical procedures

#### Hydrocarbon

Leak checks of the VOC sampling train were conducted before and after the charge was dried. A valve was closed at the probe tip and a 3-way valve was closed at the back of the analyzer. All components from just behind the probe tip to the valve at the back of the analyzer were placed under a 15-20 inHg vacuum. Less than one inHg pressure change during two minutes is acceptable and this was met.

Total flow and sample flow to the analyzer were checked using an NIST-traceable flow meter. Total flow is measured with the dilution gas off and is equal to both the sample flow from the kiln when the dilution is off and the total volume drawn by the analyzer. Sample flow is measured with dilution gas on (if used for that interval) and is the volume of gas sampled from the kiln when the dilution gas is on. This was done at the beginning and end of each sampling interval. The meter was attached to the system near the probe tip within the heated box. The valves were repositioned so that the sample came from the flow meter rather than the kiln. Readings of flow were made with the dilution gas both off and on. The flow readings were verified by observing the analyzer reading for span gas with the dilution gas off and on. The dilution ratio calculated based on the analyzer readings was always within 5% of that determined by the flow meter and almost always within 2%. Note that dilution was not actually used for runs 1 and 2 because the kiln wet-bulb was low enough initially that the gas moisture content was less than 15% until the wet-bulb temperature was greater than 130-135°F.

Calibration of the zero and span of the detector was done at the beginning of each run (about every six hours). The calibration gas was introduced by setting the valves so the calibration gas entered the system in the white heated mixing box at ambient pressure. The calibration was checked at the end of each run with no adjustments made to the instrument's zero or span during the run. A span drift less than 10% of the span value was acceptable. A zero drift of less than 3% of the span value was acceptable. A total calibration drift less than 10% was acceptable for a sampling run. These criteria were met.

#### HAPs

#### Lab analysis for aldehydes

Aldehyde standards were prepared by the dilution of neat aldehydes in water (to 400 ppm for formaldehyde, propionaldehyde, acrolein and acetaldehyde). This stock solution was mixed a BHA solution [from ortho-benzylhydroxylamine hydrochloride (BHA) and deionized water (30g BHA per liter of water)]. The stock solution-BHA mixture was vigorously agitated and allowed to sit for six hours to allow for derivatization of the aldehydes into aldoximes. The derivatized aldehyde solution was extracted with three aliquots of hexane to create a 300 ppm stock

solution in hexane. This was diluted by mass to make standards down to 0.25 ppm. 1.8 mL aliquots were place in GC autosampler vials with 10  $\mu$ L of 27,830 ppm nitrobenzene added to each as an internal standard.

The samples (from the bottles collected in field) were prepared by performing three extractions in a separatory funnel. The first extraction was with the hexane added in the field plus a 3.5 mL aliquot. The second extraction was with a 7-mL aliquot of hexane again placed in the jar with the liquid fraction. The final extraction was done with 7 mL of clean hexane in the separatory funnel after rinsing the jar with the hexane. The total hexane volume was approximately 20 mL. The volumes of the two phases were calculated from their weights. A 1.8 mL aliquot of the hexane fraction was transferred to an autosampler vial and spiked with internal standard.

The analytical instrument was a Shimadzu GC model 2010 with a flame thermionic detector (FTD), the Shimadzu equivalent of a nitrogen phosphorous detector (NPD). The column was a 105-meter Restek RTX-5 capillary with a 0.25 mm outside diameter and a stationary phase thickness of 0.25  $\mu$ m. The oven schedule was: 2 minutes at 120°C, 2°C/min ramp to 160°C, 40°C/min ramp to 220°C and 6.5 minutes at 220°C. The column flow was 25 cm/sec, with 3 mL/min septum purge, and a 1:10 split ratio with a glass wool packed split injection liner. The detector make up He was set to 20 mL/min and the H<sub>2</sub> was set to 3 mL/min. The air was set to 140 mL/min, and the source current was set to 2 pA. The He and H<sub>2</sub> gases were grade 5 and the air was grade 0.1. The injector temperature was 200°C and the detector temperature 280°C. An AOC-20i autosampler was used to perform 1  $\mu$ L injections using a 10  $\mu$ L syringe with a steel plunger.

#### Lab analysis for alcohols

Standards for methanol, phenol, ethanol, and acetic acid were prepared by the volumetric dilution of neat reagents in water. The mixed standard was prepared at a concentration of 900 milligrams per liter (mg/L). Additional standards were prepared by the volumetric dilution of the mixed standard at a range from 0.5 mg/L to 500 mg/L. Aliquots of these were placed into autosampler vials with 10  $\mu$ L of 29,455 ppm cyclohexanol internal standard.

Samples were prepared by transferring aliquots of the previously hexane-extracted aqueous fractions into autosampler vials and adding internal standard. The analytical instrument was a Shimadzu GC model 2010 with a FID detector. The column was a 60-meter Restek Stabilwax capillary with a 0.53 mm outside diameter and a stationary phase thickness of 1.5  $\mu$ m. The oven schedule was: 3 minutes at 60°C, 10°C/min ramp to 80°C, 3 minutes at 80°C, 10°C/min ramp to 230°C, and 10 minutes at 230°C. The column flow was 30 cm/sec, with 3 mL/min septum purge, and a 1:10 split ratio with a glass wool packed split injection liner. The detector make up He was set to 25 mL/min and the H<sub>2</sub> was set to 50 mL/min. The air was set to 500 mL/min. The He and H<sub>2</sub> gases were grade 5 and the air was grade 0.1. The injector temperature was 175°C and the detector temperature

250°C. An AOC-20i autosampler was used to perform 1  $\mu$ L injections using a 10  $\mu$ L syringe with a PTFE plunger.

# 8. Field data sheets and sample calculations

# Field data sheets

Samples of field data sheets are shown in Figures 10 to 13. All field data sheets are in Appendix 2 this report in electronic format (pdf).

FIE	LD DATA	SHEET FOR	TOTA	L HYDROCARBON	ANALYZER -	BEFORE	FIEL	D DATA SHEET FO	R TOTAL HYDROC	ARBON ANALYZ	ER - AFTER
BACKG	ROUND	FORMATIO	N				Operator:	ANM.	E	vent (kiin charge)	Cascade - 1
Event (k)	In charge	Castade	1	Dry-balls to	mperature	1070	Run (sampi	e) <u>3</u>	L	iboratory tempera	uae: 77 *F
Runt	3			Wel-buin t	emperature:	14-5	END TIME:	18:03			
		1 <u>RM</u>		Target Dilu	ation Ratio (TDF	n: 0.10					
Date.	<u></u>	9-15		Laboratory	temperature:	76 4	Range setti	ing on Analyzer:	er er	a Computer:	2
ANALYZI	R CALIB		(m. 2013) M.		0 L: 5=(41.4=vanit)	3-con; 2,1- off ]	Reset range	e to 3. Ranger	3		I tanks on
	Range	Analyzer, H	pading	Computer, ppm	Within range	Pot swings	34				
2010	3	0.00	Ge.	- 1260 m		495	CHECK DI	LUTION FLOW AFT	ER RUN parast conger	tid [4=vent 3	-on; 7=0H; (1,5=00)
span	3	193	(1.99)	1.98 (160)		2.79			Analyzer roading	Cons	outor, ppm
mid	3	6.50	( <b>2.1</b> 5)	0,50 mass	45 to 55		5	Spanin Ake	1.1.7	4.13	
mid	2	5.00	(k.7%)				Sample tlav	w rate (SFR)	108.5 ml	imin [3 = off, 4= o	neter (2=oft 1,5=ort)
							Read diasic	on metor:	1 set	1	
tesat ran	ge to I.	Range	3				Total flow n	ate (TFR):	<u>['761</u> mL	imin [1,5=et	i; (4×meter 2,3 ×ofi)
ETDILL	TION FL	OW BEFORE	RUN	da mi) etaman padat			Dilution rati	- Contractions and the	100A		SFR/TFR
	rate (TFF			256 mLimin IV			CHECK OF	ANALYZER CALIB			. 4=vent (1.2.5 =off)
arget dit	ution flow	rate (TDFR)		23 mLimin	[TFR x (1 - 0		[	Analyzer reading	Computer, ppm	Within range	Pot settings
et and n	and dilutio	et meter:		15 soln	[ sofh = mLAn	in * 0.0)212 ]	span	1.9%	1.44	187 10 206	2170
ampie b	ow rate (S	FR)	_1	153 mLinin	1	1 and 5 + m ]	birt	0.451	0.51	45 to 55	1. D. 49-18-1
01020-002							zero	0.01	0.00	-5 to +5	434
Spare	,			(\$0%) 15FR/TF	Diffe R   Idon(04 su	rence, 4 - OH rockOR eac	Dikition rati	o (DR <sub>Span</sub> ); o difference:	.601_ 		Spancium / Span
START T		4:43 (elo	<b>N</b>		: 1, 2,≖on; 3, 4	= off; \$=on }	End time for Conscients:		<u>146.11</u>		
ALL I LI	an FUMING	100 million	- 10	a - i-absolt - ivo; Mo	water strends,	and of 1					(1),11,11,11,11,11,11,11,11,11,11,11,11,1
NALYZE	R RANG	E ON COMP	UTER	set to match analy	reer):		Construction of the local distance				
URN OF	f Tanks										

Figure 10. Sample of field data sheet for hydrocarbon analyzer.

	DUATA SHEET, 10	5 HAPS MEASUREME	ENT	FIELD D.	ATA SHEET, 105 HA	PS MEASUREM	ENT
	TRAIN #1 - BE	FORE SAMPLE			TRAIN #1 - AFTE	R SAMPLE	
Operator: <u>Mf.M</u> Date <u>2 · 9 · 15</u>		Event (kiln charge Run (sample):		Operator: <u> <u> </u></u>		Event (kiln sturge Eun (sample)	
Antimater setting: Isopropanol rinse or la IMPINGER WEIGHTS	b wash: S.	(add 5HA to imong) Wer Weight, g	ars) BHA added, ç	Attender setting Sample fine dinse via:			
impinger #1.A	41.54	5600	(~15 mil)	al 0.45 Alles	Elapsed time.	Brie Ser	
impinger #18 Impinger #10	47.31 47.33	63.77	j- 6 ri(;	Sample flow rate : 455	sample flow rate. S	hut of other flows	using lower valves.
Optional Spike, the mercers to status rain Lab (ambient) tempera	_	oke type. Alcohol p	Adehyde c	Sample line unse val		1.18	
toir increments on tomaraie) Lab (ambient) tempera	ture <u>77</u> 4F		Company on the second	Sample line rinse vial	E2/0 mar 13	ningine samula in polises	T
ter infrees to section scale) Lab (ambient) tempera Leak check: Vacuum	ture <u>77</u> 4F <u>765</u> inHg	≇ Vacuum check	~	Sample line unse val Empty bottle weight IMPINGER WEIGHTS	Wex Weight, g		Viater removed, g
ter infrem trisin ten series Lab (ambient) tempera Leak check: Vacuum	ture <u>77</u> 4F <u>765</u> inHg	≇ Vacuum check	~	Sample line unse vial	Wer Weight, g	ningine samula in polises	Water ramoved, g
it. Sin anytingens to kills from a cairs)	ture <u>77</u> 4 <u>765</u> inHg 4568 mt/min () CLOCK Trive	≇ Vacuum check	~	Sample line inse val Emply bothe weight, IMPINGER WEIGHTS Impinger #1A	Wex Weight, g	ningine samula in polises	Viater ramovod, g

Figure 11. Sample of field data sheet for HAPs collection.

Charge:	Cascade-	1			1	Slart		ute 2015	Ti Gil	me SS							Page	11
						End			152									
Ciack	Elapsed	Rue	[		Yemp	eratures			<b></b>			HOWS				1	Victor	
bime	time	#	î dry ≅	T wet	Vave *F	G lose *F	Box *P	Chilter IC	Flow 1 Umin	Flow 2 Umin	Flow 3 L/min	Colution SOFM	Line 1 mi/min	Line 2 mirmin	Line3 mlimin	Line 1 inHG	Line 2	Line3 inHG
1.3	0.00	1	67	64	22	245	249	-1	55	Ő	0	0	45		-	270		-
38	0:30	j.	92	83	252	245	24	-/	65	0	0	0	425		—	265		
7,48	1:39	1	98	9%	25	242	242	-1	515	Ó	0	0		_	_			
1.25	3.17	11.11 B	122	INI	28	245	24	•1	-4	10	0	0					-	-
1:42	3:34	5	109	105	250	245	241	}	85	35	0	0		-	~	ſ		-
1.07	3.59	2	110	102	250	245	240	7	.79	32	15	0		-	i	-		
1:40	5.32	2	118	113	250	245	239	-1	162	63	0	0	~~		_	_		-
3118	7.16	3.5	123	120	250	245	25	-)	SI	0	C	0		456	-	_	2>	
+32	\$ 35	2	132	123	250	245	24	T	10.9	C	6	0	-			-		_
7.21	1:13	3	14		31	35	239	-1	133	0	0	15	-	-	-	-		-
1:52	(144	3	148	143	250	245	240	-	7.85	0	0	1.5	-	-	-	_	-	-
553	1243	4	150	47	250	145	240			0	12	15	<u>ش</u>		-	100	-	
1.5-1	3:5	4	150	146	2,50	245	2:0	-/	14.45	27	0	1.5			-		-	~
	4:46	4	150	45	250	145	74	-1	533	0	0	1.5		~				
54	5:64	4	50	14=5	150	245	140	-: 1	57.7	0	0	1.5		-	-	-	-	~
157	16:25	4	1550	14	193	145	540	-1	653	0	0	1.4	-	~	-	-	~	-
5:52	Milit	4	1350	165	250	143	120	1	50.0	0	0	15	-		~			-

Figure 12. Sample of kiln log data sheet.



Figure 13. Sample of flow measurement record.

#### Calculations

The "FlowCalc" worksheet in the Excel files "Kiln, Cascade.XLS" in Appendix 2 shows the calculations for each 3-minute interval during the charges. Column A is a reading number. Columns B and C are the clock and charge times, respectively. Columns D/E and F/G are the average dry- and wet-bulb temperatures.

# Humidity

Column H is the vapor pressure ( $P_{vp}$ , Pa) of water at the wet-bulb temperature. The absolute humidity (AbHum, kg<sub>water</sub>•kg<sub>air</sub>-1) is shown in column I and the molal humidity (mol<sub>water</sub>•mol<sub>air</sub>-1) in column J. These are calculated based on the dry-bulb temperature (T<sub>d</sub>, °C) and wet-bulb temperature (T<sub>w</sub> °C),

 $P_{vp.} = P_{ambient} * 10^{(16.373 - 2818.6/(Td+273.16) - 1.6908*LOG10(Td+273.16) - 1.6908*LO$ 

0.0057546\*(Td +273.16) + 0.0000040073\*(Td +273.16)\*\*2)

AbHum = (MW<sub>water</sub> / MW<sub>air</sub>) \* (1 / (P<sub>kiln</sub>/P<sub>vp</sub>-1)) - ((T<sub>d</sub>-T<sub>w</sub>) \* R<sub>psy</sub>) /  $\lambda$ 

MolHum = AbHum \* MW<sub>air</sub> / MW<sub>water</sub>

where MW are molecular weights (kg•kgmol<sup>-1</sup>), R<sub>psy</sub> is the psychrometric ratio (0.95 kJ•kg<sup>-1</sup>•K<sup>-1</sup>), and  $\lambda$  is the latent heat (2419 kJ•kg<sup>-1</sup>).

### Flows

The volumetric dry gas flow rate (DryGasV, L•min<sup>-1</sup>) in column K is the flowmeter reading adjusted for the meter calibrations and the molar humidity of the entering gas. This is in standard (at 0°C) liters per minute. In column L this has been converted to a mass flow rate (DryGasM, kg•min<sup>-1</sup>) and in column M is the same

information is expressed as a molal flow rate (DryGas, kgmol•min<sup>-1</sup>). These values are for the dry gas vented from the kiln.

DryGasV = (FlowMeter1 + FlowMeter2 + FlowMeter3) \* (1/(1+MolHumin))

 $DryGasM = (DryGasV L \cdot min^{-1}) * 1/(22.4 m^{3} \cdot kgmol^{-1}) * MWair / (1000 L \cdot m^{-3})$ 

DryGas (kgmol/min) = DryGasM / MWair

The water removal rate (WaterVented, g•min<sup>-1</sup>) (column N) is calculated from the humidity (column I) and the gas flow (column L). The total water (column O) is an integration of column N over time.

WaterVented = (AbHum - AbHum<sub>In</sub>) \* (DryGasM \* 1000 g•kg<sup>-1</sup>)

#### Moisture content

The moisture content of the wood at each three-minute interval (column P) was determined by reducing the moisture content of the wood from the previous value by accounting for the amount of water leaving the kiln during the interval.

 $MC = MC_{Previous} - 100 * (WaterVented / (1000 g \cdot kg^{-1}) / ODWoodWt)$ 

This amount is then adjusted by adjusting the wet-bulb temperature to make the ending moisture content match that measured by ASTM D4222.

## Hydrocarbon

The original total hydrocarbon analyzer reading is shown in column Q. In column R this has been corrected to compensate for the range setting switch on the analyzer. Also in column R, the THA data between sampling runs (rows labeled "test" in column AA) has been adjusted to the average of the data during the 9-minute period before and the 9-minute period after the analyzer testing and calibration time.

The dilution THA (column S) is the corrected THA reading divided by the dilution ratio (from column AA). In column T we have the opportunity to compensate for the effect of moisture on the JUM detector. Column T equals column S because dilution was used and no compensation was made. Finally in column U, the hydrocarbon concentration is converted to a dry gas basis concentration using the molar humidity (column J).

THC<sub>Dry</sub>, ppm= THC \* (1 + MolHum)

In column V, the hydrocarbon flow rate (THC<sub>Vented</sub>, g<sub>Carbon</sub>•min<sup>-1</sup>) is calculated in a manner analogous to the water flow rate using the dry gas flow rate and the hydrocarbon concentration.

 $THC_{Vented} = DryGas * (THC_{Dry} / 10^{6}) * MW_{Propane} * (1000 \text{ g} \cdot \text{kg}^{-1}) * (0.81818 \text{ g}_{Carbon} \cdot \text{g}_{Propane}^{-1})$ 

Column W is the integral of column V over time, the cumulative hydrocarbon released up to that point in the schedule (in grams). Column X is the cumulative unit emissions, that is, column W divided by the oven-dry weight of the wood in the kiln. Column AI is the cumulative emissions in pounds per thousand board feet and column AH is the rate of emissions release (lb•mbf<sup>-1</sup>•hr<sup>1</sup>)

Column Z indicates the hydrocarbon sampling run and column AA is the dilution ratio during that run.

The remaining columns are used not used in the hydrocarbon calculations. They are for graphing shown on other worksheets in the workbook.

At the end of the FlowCalc spreadsheet (at the bottom) are summaries by run of the flow data for the total hydrocarbon run intervals (interval summary button will reposition spreadsheet).

Moisture content and board weight data are on the "Define" worksheet and the original data are in the files named "Weights, Cascade.XLS".

#### HAPs

Within the file "HAPs, Cascade.xls", the summary page presents the data by run interval. The data is copied from the other pages to provide a concise summary.

The "Field Data" page is data transcribed from the field data sheets (copies of the sheets are included in Appendix 2 in PDF format) and includes the ambient pressure, lab temperature, flow rate through the impingers, and run start and stop times.

The "Laboratory Data" page contains results from the lab analysis for HAPs. These values come from the files "AQU, GC Sheet, Cascade.xls" and "ALD, GC Sheet, Cacade.xls" in the "Lab Data" directory. The GC retention times and peak areas and the GC calibrations are in these files.

On the "Impinger Calculations" page, the field data and laboratory data are used to give a dry gas flow rate through the impingers (columns J and K) and the mass of target compounds in the impingers (columns L to Q). Flow rates were adjusted to standard conditions in columns F and G.

ImpgrFlowstd\_mL = ImpgrFlow \* (273.16K / Tmeter) / (Pmeter / 101.33 kPa)

A dry gas flow rate is calculated in columns H and I

 $ImpgrFlow_{Dry_mL} = ImpgrFlow_{Std_mL} / (1 + MolHum)$ 

The average of the before and after gas flow measurements through the impingers (column J) is then converted to a mass basis in column K.

ImpgrFlow<sub>Dry\_g</sub> = MWair\* ImpgrFlw<sub>Dry\_mL</sub>\* P / (T \* R)

Finally, the mass of each compound recovered from the impinger is calculated in columns L to S.

## Mass<sub>i</sub> = (Concentration<sub>i</sub>) / (DenSolvent) \* (Mass solvent)

The "Kiln Calculations" page uses a ratio of the dry gas flow through the kiln (calculated in the spreadsheets named "Kiln, Cascade.xls" and copied to column D) to the dry gas flow rate through the impinger to scale up the quantities and obtain the mass of each compound leaving the kiln (columns I to P).

On the "Emission" page, the amount of a HAP leaving the kiln is divided by the mass (in kg) or volume of wood (in mbf) to express the emissions on a per kg of wood (columns B-I) or per mbf basis (columns J-Q). Concentrations leaving the kiln are given in columns R to AG.

The "Quality Assurance" page presents information on the spikes, duplicates and blanks. For each spike a % recovery is calculated based on the mass of a HAP recovered divided by the amount added. The difference for each duplicate is calculated as a percentage from the difference between the impingers divided by the average mass collected after adjusting for impinger flow.

The remaining pages in "HAPs, Cascade.xls" are for graphing purposes.

# 9. Chain of custody information

Wood was collected by mill personnel and delivered to Oregon State by Cascade Hardwoods. The wood was retained by Oregon State after delivery as documented in section 1. Field samples remained at Oregon State University.

# 10. Calibration documentation

	S	ensidyne	, LP	
	CA	LIBRATION CER	TIFICATE	
Ce	I S/N: 1302015-S		Date: Feb	oruary 07, 2013
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Figure 14. Flow meter calibration.

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Figure 15. Certificates for calibration gases.

# 11. Anomalies

There were no anomalies.

# 12. Statement of validity

The statements in this report accurately represent the testing that occurred.

Michael R Milota

Michael R. Milota

Oregon Wood Innovation Center Department of Wood Science and Engineering 136 Richardson Hall Oregon State University Corvallis, OR 97331-5751 (541) 737-4210 V (541) 737-3385 F

### Appendix 1. Detailed sampling procedures

# Checks of kiln to record on log

**Purpose:** Ensure kiln is operating correctly

Clock time: Record from computer

**Run time:** Record from computer. Check the box if the computer screen being refreshed and time is advancing.

**Box temperature:** Read from plastic electrical enclosure on wall or on computer screen. The top and bottom numbers on controller should be similar and greater than the kiln temperature, 240°F.

**Line temperature:** Read from plastic electrical enclosure on wall or on computer screen. The top and bottom numbers on controller should be similar and greater than the box temperature, 245°F.

**Valve temperature:** Read from plastic electrical enclosure on wall or on computer screen. The top and bottom numbers on controller should be similar and greater than the line temperature, 250°F.

**Dry-bulb temperature:** Read from computer screen. Compare to paper graph to be sure it's correct. If it's not within a degree or two of the chart, check again in a few minutes. During startup (the first 3 or so hours), it may not be able to track. If it's too high, the heat valve should be closed, too low and the heat valve should be open. If it does not appear to be working correctly, call Mike.

**Wet-bulb temperature:** Read from computer screen. Compare to graph to be sure it's correct.

If the wet-bulb is too low, it means that the kiln atmosphere is too dry. Check the flow meters. If Flow1 is about 6 L/min (its lower limit), make sure that Flow2 and Flow3 are turned off. Flow2 records automatically. Enter any Flow3 change into the computer. Otherwise, call Mike.

If it's too high, then either the kiln atmosphere is too humid or the sock is not being wetted. If Flow 1 is near 200 L/min (its upper limit) add venting by opening Flow2 and/or Flow 3. Enter any Flow3 change into the computer. The maximum for Flow2 is 50 L/min, if it reads over this value for several readings, reduce it to about 45 L/min. Don't change Flow3 often, rather set it and leave it for several hours if possible. Keep the Flow 3 reading constant by small adjustments. As Flow1 decreases or Flow2 turned down, there is more pressure behind Flow3 and the flow increased. Check for water in the wet-bulb reservoir (push the float down and make sure it's getting water).

Check both Wet-bulb1 and Wet-bulb2 and make sure they are reading about the same. If they differ by more than 2°F, call Mike
If both wet-bulbs are reading the same as the dry-bulb, check the wet-bulb water.

If these procedures do not correct the wet-bulb temperature within 30 minutes, call Mike.

**Chiller temperature:** Read the chiller temperature. It should be about -1°C.

**Flow 1:** Read from computer. The value of Flow1 changes depending on the wetbulb. If Flow 1 is 6 L/min and the wet-bulb is too low, there's probably nothing we can do. If it's 200 L/min and the wet-bulb is too high, Flow2 and/or Flow3 can be opened. Flow2 and Flow3 should be adjusted so that Flow1 stays below 175 to 200 L/min.

**Flow 2:** Read from computer. The value of Flow2 is set by you. It will vary a little - as flow 1 goes down, flow 2 will go up. Do not set it to > 40 L/min if you think Flow1 is going to decrease or it will go off scale and not be read by the computer

**Flow 3:** Read from meter. The value of Flow3 is set by you. It will vary a little - as flow 1 goes down, flow 2 will go up. Be sure to clearly record this value and when you change it. Change it on the computer screen (click on it and type the new value).

**Dilution flow:** Read dilution flow meter. It should read the same setting as the red flag. Do not adjust. If significantly different, investigate.

Impinger flows: Read from rotometers. This should be about 250 to 500 cc/min.

**Line vacuum:** Read from the vacuum gauge. This should be about 20"Hg.

#### Total hydrocarbon analyzer

# BACKGROUND INFORMATION

Get the dry- and wet-bulb temperatures from the kiln schedule or off the computer. Use the highest expected values for the next three to six hours.

Read absolute humidity off the psychrometric chart or table. Calculate or read from tables -

> Percent moisture = 100 / [1 + 1 / 1.61\*AbHum] Target Dilution Ratio (TDR) = 15 / Percent Moisture

Event = the name of the drying cycle. Run = the number of the 3-hour interval. Operator, that's you. Date – use date VOC run will start if close to midnight

# AMBIENT DATA

Read the laboratory temperature from the computer or thermometer.

#### ANALYZER CALIBRATION (BEFORE SIDE OF SHEET)

Set valves so that 1, 2 = OFF; 3=ON; 4=VENT. This allows gas to flow out of the vents from the calibration tanks and shuts off all other sources. Only calibration gas should go through the detector.

Open the zero gas tank valve set analyzer to range 3 zero valve on, others off set flow to 3 L/min using regulator on tank wait for a stable reading (about 30 to 60 seconds) use the zero dial (pot) on THA to get a zero reading read the analyzer read computer note pot setting Close valve on zero gas tank

Open span gas tank valve (may be 197 or 794 ppm gas) span valve on, others off set flow to 3 L/min using regulator on tank wait for a stable reading (about 30 to 60 seconds) use the span dial (pot) on THA to get a reading of 610ppm read the analyzer and record, eg, record 7.96 read computer (should read about 794) record pot setting Leave span tank valve open

Open mid gas tank valve (197 or 50 ppm gas) mid valve right on, others off set flow to 3 L/min using regulator on tank wait for a stable reading (about 30 to 60 seconds) read and record analyzer and computer (do not adjust pot settings) check for within tolerance switch analyzer to range 2 read analyzer and computer check for within tolerance switch analyzer back to range 3 Turn off mid gas tank valve

# SET DILUTION FLOW BEFORE RUN (BEFORE SIDE OF SHEET)

Set valves so that 1, 2, 3 = OFF; 4=meter. This allows gas to flow only from the meter to the detector.

Use the Gilibrator to take 4 readings of the total flow rate (TFR). This is the total flow drawn by the analyzer and should be about 1.6 L/min Make sure the average does not include any "bad" readings Record the average in mL/min; It should be 1500-1600 mL/min Write the Run # and "Pre-TFR" on the Gilibrator printout.

Calculate the next two values -Target dilution flow rate (TDFR) is the TFR x (1 - DR) Target sample flow rate (TSFR) is the TFR x DR Check that the sum of these is the Total Flow Rate

Set dilution flow Set red pointer to desired dilution flow Slowly open lower valve on dilution flow meter (1=ON) Use upper valve on dilution flow meter to adjust flow Do not adjust this meter after this point Read the meter that you just set and record the value in SCFH Calculate and record L/min

Use the Gilibrator to take 4 readings of the sample flow rate (SFR). This is the flow through the analyzer after dilution is set. It will vary, depending on the dilution setting.

Make sure the average does not include any "bad" readings Record the average in mL/min

Write "Pre-SFR" on the Gilibrator printout.

# CHECK DILUTION FLOW BEFORE RUN (BEFORE SIDE OF SHEET)

Set valves so that 1, 3 = ON; 2=OFF; 4=VENT. This allows gas to flow out of the vent from the calibration tank and shuts off all other sources. Calibration gas and dilution air will go through the detector.

Open span gas tank valve (should already be open) span panel valve right (on), others down (off) set flow to 3 L/min using regulator on tank set analyzer to range 3 wait for a stable reading (about 30 to 60 seconds), record turn off all calibration gas tank valves all calibration gas panel valves off

All tank valves off

Calculate the dilution ratio based on gas flow by dividing the Sample Flow Rate by the Total Flow Rate. DR = Absolute value of [ 100\*(DR Span - DR Flow)/DR Flow ]

Calculate the dilution ratio based on span gas by dividing the diluted span by the undiluted span.

If the dilution ratio calculated from the span gas and the dilution ration calculated from the flow do not agree within 5% - DO NOT PROCEED\*\*\*\*. Check the calculations, then redo the measurements.

\*\*\*\* check calculations, check that values for ppm and flows make sense, remeasure everything. If it still does not agree, call Mike

## START RUN (BOTTOM OF BEFORE SIDE OF SHEET)

Set valve so that 1, 2, 5 = on; 3, 4=off; all calibration tank valves off

Record the start time. Use the computer clock or stopwatch time.

Make sure analyzer is on appropriate range, usually range 3, to keep THC reading on computer between 60 and 600.

Monitor system, as needed. Record system condition at least hourly.

End time should be no more than 3-6 hours from start time.

## POST-SAMPLE PROCEDURE - AT END OF RUN (AFTER SIDE OF SHEET)

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Record your name as the operator.

Event = the drying cycle. Run = number of the 3-hour interval. Operator, that's you.

# AMBIENT DATA

Read the laboratory temperature from the thermometer.

Fill out appropriate information on Pre-sample side of data sheet for next run. This will save time in between runs.

### END TIME

Record computer time.

DO NOT adjust dilution gas or analyzer pots until the instructions tell you to.

### CHECK DILUTION FLOW AFTER RUN (AFTER SIDE OF SHEET)

Measure diluted span gas: Set valves so that 1, 3 = on; 2=off; 4=vent. This allows gas to flow out of the vent from the calibration tank and shuts off all other sources. Calibration gas and dilution air will go through the detector.

Open span gas tank valve

span panel valve ON, others OFF set flow to 3 L/min using regulator on tank set analyzer to range 3 wait for a stable reading (about 30 -60 seconds) record

Sample flow rate: Set values so that 1=on; 2, 3 = off; 4=meter. This allows gas to flow only from the meter and the dilution to the detector.

Use the Gilibrator to take 4 readings of the sample flow rate (SFR). This is the flow through the analyzer with dilution on. Make sure the average does not include any "bad" readings Record the average in L/min Write Run # and "Post-SFR" on the Gilibrator printout.

Read dilution flow meter To calculate the L/min, divide scfh by 2.12 Turn off dilution flow meter using valve 1 (lower dilution valve)

Total flow rate. Set valves so that 1, 2, 3 = off; 4=meter. This allows gas to flow only from the meter to the detector.

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Use the Gilibrator to take 4 readings of the total flow rate (TFR). This is the total flow drawn by the analyzer and should be about 1.6 L/min Make sure the average does not include any "bad" readings Record the average Write Run # and "Post-TFR" on the Gilibrator printout.

Calculate the dilution ratio based on gas flow by dividing the Sample Flow Rate by the Total Flow Rate.

# CHECK CALIBRATION OF ANALYZER (AFTER SIDE OF SHEET)

Set valves so that 1, 2 = off; 3=on; 4=vent. This allows gas to flow out of the vents from the calibration tanks and shuts off all other sources. Only calibration gas should go through the detector.

Span gas tank valve should be open

span panel valve ON, others down OFF set flow to 3 L/min using regulator on tank set analyzer to range 3 wait for a stable reading (about 30 -60 seconds) read analyzer (do not adjust pot settings), record, for example, 7.94 as 794 read computer (should read about the same) note pot setting check for within tolerance

#### Open mid gas tank valve

mid panel valve = ON, others OFF set flow to 3 L/min using regulator on tank set analyzer to range 3 wait for a stable reading (about 30 -60 seconds) read analyzer (do not adjust pot settings), record, for example, 1.97 as 197 read computer (should read same as analyzer) check for within tolerance

### Open the zero gas tank valve

zero panel valve = ON, others OFF set flow to 3 L/min using regulator on tank wait for a stable reading (about 30 -60 seconds) read analyzer (do not adjust pot settings) read computer note pot setting

Close all tank valves if charge is ending

Calculate the dilution ratio based on gas concentration by dividing the Diluted span by the Span

Calculate % difference in the two dilution ratios as 100 \* {Absolute Value (DRSpan-DRFlow)} / DRFlow

Record the time now as the end time for check.

Start Pre-Sample procedure for next run.

# HAP 105 Collection

# BACKGROUND DATA

Begin about 15 minutes before run should start Operator, that's you. Date, today or tomorrow if sample will start after midnight Event = Kiln Charge Run = sequence of M/F measurement (1-A, or 5-C, etc)

## PRE RUN DATA

Call 9-541-754-0081 and get altimeter setting.

#### **IMPINGER WEIGHTS**

Verify that the impinger weights match the prerecorded weights on the data sheet.

Put 15 mL of BHA solution in impinger #1. Put 10 mL of BHA solution in impinger #2. Impinger #3 is not filled. It is for overflow.

Reweigh the impingers with the BHA solution. Place BHA stock back into cooler Install impingers and lower into chiller

# LEAK CHECK

Read the laboratory temperature. Close valve to sample probe. Turn on pump (it may already be on) Evacuate to 15 to 18 " Hg, record Close valve that is near pump Note pressure and start timer Allowable pressure change is 1" Hg in 2 minutes, if it is more than this, find the source of the leak. Record change. Slowly open valve near probe tip so that pressure is slowly relieved. Completely open valve near probe tip Open valve near pump

#### SAMPLE FLOW RATE

Attach probe tip to Gilibrator Take 4 readings Make sure all readings in average are "good" readings Record the average

## START TIME

Put probe into kiln (or turn valve to sample kiln) and record time. Check meters to make sure gas is flowing

## FLOW READINGS DURING TEST

Note flow meter reading at intervals of at least 20-30 minutes Run test for 1:30 or less if impingers fill

# POST RUN DATA

Begin about 10 minutes before run should end Label a sample bottle with the Event and Run numbers and record the weight. Call 9-541-754-0081 and get altimeter setting.

#### END TIME

Remove probe (or turn valve to meter setting) from kiln Record time

## SAMPLE FLOW RATE

Rinse probe with 5 mL of DI water Read the laboratory. Attach probe tip to Gilibrator Take 5 readings Make sure all readings in average are "good" readings Record the average

#### **IMPINGER WEIGHTS**

Lift impingers from chiller, take to scale, and place onto rack Dry the outside of the impingers Remove U tubes connecting the impingers together Weigh sample bottle with lid Weigh the impingers (without stoppers) with the catch and record Transfer the impinger contents to the sample bottle Weigh the sample bottle with lid and record Rinse impingers (last to first) with 10 mL DIW (save in the sample bottle) Weigh the sample bottle with lid and record Rinser impingers (last to first) with 5 mL hexane (save in the sample bottle) Weigh the sample bottle with lid and record Rinser impingers (last to first) with 5 mL hexane (save in the sample bottle) Weigh the sample bottle with lid and record Rinser the sample bottle into cold storage Record the volume of any liquids lost during this procedure. Wash glassware with phosphate-free detergent and set out to dry. Appendix 2. Electronic copy of data and calculations

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