Total hydrocarbon and HAP emissions from the drying of Douglas-fir lumber

Report to

Columbia Vista Corp. 18637 SE Evergreen Hwy Vancouver, WA 98684 Phone: 360-892-0770 Contact: Devin Sanders, DSanders@ColumbiaVista.com. Air Discharge Permit 09-2854

Uri _____ Randy _____ Wess _____ Clint _____ John _____ Vannessa _____ Jerry ____ Brian _____ Brian _____ Duane _____ Duane _____ Allison _____ Chip _____ Traci _____ Traci _____ Tile _____

Report by

Michael R. Milota Department of Wood Science and Engineering Oregon State University Corvallis, OR 97331 541-737-4210 Mike.Milota@OregonState.edu

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Summary

One charge of Douglas-fir 4x5 lumber was dried in a small kiln at Oregon State University. The kiln dry- and wet-bulb temperatures were based on a schedule provided by Columbia Vista. The maximum temperature was 175°F (79°C). The air velocity was 750 feet per minute (3.7 m/s). The kiln was indirectly heated with steam. The amount of air entering the kiln was regulated to control humidity.

A JUM VE-7 total hydrocarbon analyzer was used to measure organic emissions following EPA Method 25A. The results are shown in Table 1.

Table 1. Summary of total hydrocarbon results to 8% moisture content. VOC units are pounds per thousand board feet as carbon.

Charge	Initial MC	Final MC	Time to 8%	VOC ^B
	%	%	hr:min	lb/mbf
Douglas-fir	39.5	15.0 ^A	143:28	0.86

^A actual time to 14.3% MC was 150:00 hours

^B VOCs are reported at 15% moisture content

NCASI Method ISS/FP-A105.01 was used to measure the MACT HAP emissions. The results are shown in Table 2. The sum of the HAPs emitted was 0.13 lb/mbf for the Douglas-fir.

Table 2. Summary of HAP results for moisture content and time shown in Table1. Emissions units are pounds per thousand board feet.

	Methanol	Phenol	Form- aldehyde	Acet- aldehyde	Propion- aldehyde	Acrolein
Douglas- fir	0.084	0.0	0.0016	0.042	0.0002	0.0008

^A None detected

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1. Description of source

The tested source is a lumber dry kiln. Lumber destined for the mill's kiln was sampled and tested in a small-scale kiln at Oregon State University.

Mill personnel reported that the log source was forests in the Woodland, Washington area. The trees were inland and 80-90 years of age. They were harvested within 30 days of the sawing date. During that time they were stored on land. The Douglas-fir was sawn at Columbia vista on March 3, 2015.

Enough 4-foot pieces of wood for three charges of lumber were delivered to Oregon State by an employee of Columbia Vista on March 5, 2015. The wood was palletized and wrapped in plastic at the mill to prevent moisture loss during transport. The wood appeared to be fresh.

At OSU, the 4-foot pieces were sorted on March 6. One third was randomly pulled from the pile and used for the charge. It was wrapped and stored in a cooler at 40°F. The remaining wood was wrapped in plastic and placed in a freezer at 10°F.

2. Date and time of test

The charge was dried from Monday, March 9 at 6 am until noon on Sunday, March 15, 2015. Drying was done under the supervision of Mike Milota at Oregon State University. Students were used to monitor parts of the test.

3. Results

Total hydrocarbon

See Table 1, page 1, for a summary of the hydrocarbon results. Details for each sampling interval are tabulated (Table 3) and the hydrocarbon emissions are summarized graphically (Figures 1-3) here. All emission data is presented in detail in electronic form in Appendix 2.

An interval is the period between analyzer calibrations, about six hours of data. The interval time periods shown in the table include the calibration times and mass calculations are adjusted to account for these. Sampling occurred for approximately 95% of the drying time.

Figure 1 shows total hydrocarbon concentration (left scale) and dry gas vent rate (right scale) versus time. Concentration has a peak at 24. In general, the concentration then decreases though the schedule. Once the drying rate slows (this was thick wood), the kiln venting remains nearly constant and at a minimum. The concentration then deceases as the drying rate decreases. The vent rate is quite variable until 40 hours due to heating of the wood.

Figure 2 shows the cumulative hydrocarbon emissions (left scale, smooth line) and the rate of emissions (right scale, jagged line) versus time. The cumulative value is the emissions up to any point in time in the schedule. The rate is how much is coming out per unit time. The maximum emission rates occurred between 24 and 72 hours, after which it steadily decreases as the moisture loss from the wood slows.

Figure 3 shows the total hydrocarbon emissions as a function of wood moisture content. This graph would be useful for predicting emissions at various final moisture content levels. As is typical, the hydrocarbon release rate is linear with moisture loss at the lower moisture content.

Sample	Time	Cumu	lative	Average	Flow	rate	THC mass	THC con	centration	THC rate	THC mass	THC rate		Average	
Run		Dry Gas	Water	Humidity	Dry @68	Wet @68	as C	wet	dry	as C	as C	as C	Wood MC	Air MC	Anal. MC
	hrs	kg	kg	kg/kg	I/min	1/min	9	ppmv	ppmv	g/hr	lbs/mbf	lb/hr/mbf	%	%	%
1	6.06	8,93	0.16	0.018	20.4	21.0	0.31	36.1	37.4	0.05	0.005	0.0009	39.4	2.8	2.8
2	6.06	2.99	0.15	0.051	6.8	7.4	0.53	130.9	142.3	0.09	0.009	0.0015	39.3	7.5	7.5
3	6.26	3.07	0.31	0.100	6.8	7.9	1.16	259.7	302.1	0.18	0.020	0.0032	39.0	13.9	8.2
4	5.66	6.23	1.04	0.167	15.2	19.3	3.57	347.0	436.3	0.63	0.061	0.0109	38.6	21.2	12.5
5	5.91	9.53	1.74	0.183	22.3	28.9	6.56	447.3	579.9	1.11	0.113	0.0192	36.9	22.7	13.6
6	6.21	12.30	2.47	0.200	27.4	36.2	6.99	356.5	472.2	1.13	0.121	0.0194	34.8	24.4	15.4
7	6.01	9.34	2.07	0.221	21.5	29.1	4.59	293.0	397.8	0.76	0.079	0.0132	32.7	26.3	12.9
8	5.91	7.40	1.79	0.242	17.3	24.1	3.68	290.3	403.4	0.62	0.064	0.0108	30.8	28.0	13.7
9	6.01	9.21	2.26	0.246	21.2	29.6	3.45	216.4	302.1	0.57	0.060	0.0099	28.7	28.4	13.9
10	5.96	7.56	1.86	0.246	17.5	24.5	2.66	202.6	282.9	0.45	0.046	0.0077	26.6	28.4	13.9
11	6.01	4.44	1.09	0.246	10.2	14.3	1.58	206.7	288.6	0.26	0.027	0.0045	25.1	28.4	11.6
12	6.01	2.45	0.59	0.240	5.6	7.8	0.89	210.4	292.2	0.15	0.015	0.0025	24.4	27.9	11.4
13	7.06	5.47	1.25	0.228	10.7	14.7	1.77	190.3	260.4	0.25	0.030	0.0043	23.4	26.9	11.0
14	5.01	3.82	0.80	0.210	10.6	14.1	1.13	177.1	237.0	0.22	0.019	0.0039	22.4	25.3	10.4
15	6.06	4.44	0.91	0.206	10.1	13.5	1.25	170.7	227.4	0.21	0.022	0.0036	21.5	24.9	11.0
16	5.96	4.36	0.82	0.187	10.1	13.2	1.19	168.9	220.0	0.20	0.021	0.0034	20.7	23.2	10.2
17	6.06	4.57	0.76	0.167	10.4	13.2	1.16	161.4	204.7	0.19	0.020	0.0033	19.9	21.2	9.1
18	4.96	3.96	0.63	0.158	11.0	13.8	0.97	156.8	197.0	0.19	0.017	0.0034	19.2	20.3	9.5
19	6.96	5.10	0.65	0.127	10.1	12.2	0.72	94.8	114.3	0.10	0.012	0.0018	18.6	17.0	7.6
20	5.91	4.38	0.65	0.149	10.3	12.7	1.04	154.2	191.2	0.18	0.018	0.0030	17.9	19.4	9.9
21	6.11	4.57	0.69	0.152	10.3	12.9	1.14	161.3	200.9	0.19	0.020	0.0032	17.3	19.7	10.0
22	5.81	4.45	0.66	0.149	10.6	13.1	1.04	152.4	189.2	0.18	0.018	0.0031	16.6	19.4	10.7
23	6.01	4.62	0.70	0.151	10.6	13.2	1.17	163.9	203.8	0.19	0.020	0.0034	15.9	19.6	11.0
24	5.51	4.21	0.62	0.147	10.6	13.1	1.03	159.3	196.9	0.19	0.018	0.0032	15.3	19.1	10.7
Sum	143.45	137.4	24.7				49.6			8.304	0.855	and the second second		360.2	
Average				0.175	13.2	17.1		204.5	265.8	0.3460		0.0060			

Table 3. Summary of results for each sampling interval for total hydrocarbon.



Figure 1. Hydrocarbon concentration and vent rate versus time



Figure 2. Cumulative hydrocarbon emissions (left scale, smooth line) and the rate of emissions (right scale, jagged line) versus time.



Figure 3. Total hydrocarbon emissions as a function of wood moisture content.

HAPs

See Table 2, page 1, for a summary of the HAP results. Details for each sampling interval are tabulated (Table 4) and the HAP emissions are summarized graphically (Figures 4 and 5) here. All emission data is presented in detail in electronic form in Appendix 2.

A summary of the kiln conditions for each sampling interval is shown in Table 4. A collection interval is the time the impingers were on and sampling occurred, approximately 90 minutes. An adjusted interval is the period spanning the midpoints between collection intervals, about six hours. For example, if the impingers were on from 12:00 to 13:30, 18:00 to 19:30, and 00:00 to 1:30, the 18:00 to 19:30 impinger set represents the adjusted interval from 15:45 to 21:45. The mass calculations are adjusted proportionally to represent emissions during the adjusted interval. For example, if a collection interval was 90 minutes and the adjusted interval was six hours, the amount of HAPs in the impinger is multiplied by four. Sampling occurred for 26.7% of the drying time.

The MACT HAP emissions and the emissions of ethanol and acetic acid are shown in Table 5. The total HAP emissions were 0.13 lb/mbf for the Douglas-fir (does not include the non-HAPs, ethanol and acetic acid). Among the HAPs, methanol was emitted in the greatest quantity, 0.084 lb/mbf and acetaldehyde was about half of this, 0.042 lb/mbf. Other HAPs (formaldehyde, propionaldehyde, and acrolein)

are present and comprise only 1.9% of all the HAPs. Phenol was not detected in any sample.

The HAP emissions as a function of time and wood moisture content during the cycle are shown in Figures 4 and 5, respectively. The rate of HAP emissions generally decreases with time throughout the schedule (lines are mostly concave downward in Figure 4) for most HAPs. The rate of HAP emissions per percent moisture content change are mostly linear with moisture content (straight lines in Figure 5) except for a decreasing release for acetaldehyde.

	Collection	Adjusted	Dry gas	Average	Molar	Mois	ture
Sample	Interval	Interval	mass	Dry gas	Humidity	Con	tent
Run ID	8			flow rate		Mid	End
	hours	hours	kg	kg/min	mol/mol	%	%
1	1.50	3.76	6.782	0.030	0.027	39.4	39.4
2	2.28	6.01	3.980	0.011	0.057	39.3	39.2
3	1.50	6.01	2.944	0.008	0.127	39.1	39.0
4	1.50	5.96	2.973	0.008	0.216	38.8	38.6
5	1.52	6.01	11.166	0.031	0.286	37.6	36.7
6	1.50	6.01	11.234	0.031	0.314	35.7	34.5
7	1.48	6.01	9.880	0.027	0.344	33.5	32.4
8	1.50	6.01	7.670	0.021	0.378	31.5	30.6
9	1.50	6.01	8.965	0.025	0.396	29.6	28.4
10	1.50	5.96	8.285	0.023	0.396	27.3	26.4
11	1.50	6.01	6.185	0.017	0.396	25.5	24.9
12	1.50	6.01	2.242	0.006	0.394	24.6	24.3
13	1.50	6.01	4.037	0.011	0.375	23.9	23.4
14	1.50	6.01	4.642	0.013	0.356	22.8	22.3
15	1.50	6.01	4.454	0.012	0.333	21.9	21.4
16	1.52	5.96	4.353	0.012	0.321	21.0	20.6
17	1.50	6.01	4.466	0.012	0.276	20.2	19.8
18	1.50	4.76	3.697	0.013	0.263	19.4	19.2
19	1.50	5.21	4.033	0.013	0.226	19.0	18.7
20	1.50	7.26	5.303	0.012	0.219	18.4	18.0
21	1.53	6.76	5.037	0.012	0.243	17.5	17.2
22	1.50	5.96	4.530	0.013	0.239	16.9	16.5
23	1.50	6.01	4.623	0.013	0.243	16.2	15.9
24	1.50	6.01	4.599	0.013	0.237	15.5	15.2
25	1.50	1.75	1.335	0.013	0.235	14.9	15.0
SUM	38.33	143.45					

 Table 4.
 Summary of HAP sampling intervals.

	Interval	Wood				Unit mass	leaving kiln			
Sample Run ID	Endpoint	Moisture Content	Methanol	Phenol	Ethanol	Acetic acid	Form- aldehyde	Acet- aldehyde	Propion- aldehyde	Acrolein
	hours	%	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf
1	3.76	39.4	0.0002	0.0000	0.0004	0.0007	0.00000	0.0007	0.00000	0.00000
2	9.76	39.2	0.0004	0.0000	0.0008	0.0010	0.00000	0.0014	0.00000	0.00000
3	15.77	39.0	0.0005	0.0000	0.0011	0.0019	0.00000	0.0017	0.00000	0.00000
4	21.73	38.6	0.0011	0.0000	0.0015	0.0044	0.00001	0.0028	0.00000	0.00001
5	27.74	36.7	0.0037	0.0000	0.0036	0.0208	0.00006	0.0065	0.00001	0.00006
6	33.75	34.5	0.0044	0.0000	0.0030	0.0220	0.00007	0.0037	0.00001	0.00006
7	39.75	32.4	0.0049	0.0000	0.0029	0.0228	0.00009	0.0039	0.00001	0.00007
8	45.76	30.6	0.0046	0.0000	0.0025	0.0196	0.00010	0.0034	0.00001	0.00008
9	51.77	28.4	0.0060	0.0000	0.0022	0.0273	0.00013	0.0032	0.00002	0.00008
10	57.73	26.4	0.0066	0.0000	0.0019	0.0238	0.00011	0.0022	0.00001	0.00005
11	63.74	24.9	0.0052	0.0000	0.0010	0.0160	0.00009	0.0018	0.00001	0.00005
12	69.74	24.3	0.0016	0.0000	0.0003	0.0053	0.00004	0.0008	0.00000	0.00002
13	75.75	23.4	0.0039	0.0000	0.0005	0.0095	0.00007	0.0011	0.00001	0.00003
14	81.76	22.3	0.0043	0.0000	0.0004	0.0101	0.00007	0.0010	0.00001	0.00003
15	87.77	21.4	0.0036	0.0000	0.0003	0.0088	0.00008	0.0010	0.00001	0.00003
16	93.73	20.6	0.0039	0.0000	0.0002	0.0114	0.00008	0.0008	0.00001	0.00002
17	99.73	19.8	0.0035	0.0000	0.0001	0.0065	0.00007	0.0008	0.00001	0.00002
18	104.49	19.2	0.0024	0.0000	0.0001	0.0044	0.00004	0.0005	0.00000	0.00001
19	109.70	18.7	0.0035	0.0000	0.0001	0.0052	0.00007	0.0007	0.00001	0.00002
20	116.96	18.0	0.0024	0.0000	0.0001	0.0023	0.00005	0.0006	0.00001	0.00002
21	123.72	17.2	0.0040	0.0000	0.0000	0.0040	0.00006	0.0006	0.00001	0.00002
22	129.68	16.5	0.00384	0.00000	0.00012	0.00540	0.00010	0.00073	0.00001	0.00003
23	135.69	15.9	0.00369	0.00000	0.00010	0.00362	0.00007	0.00070	0.00001	0.00003
24	141.70	15.2	0.00467	0.00000	0.00010	0.00422	0.00009	0.00075	0.00001	0.00003
25	143.45	15.0	0.00140	0.00000	0.00004	0.00123	0.00003	0.00021	0.00000	0.00001
		Sums:	0.084	0.000	0.023	0.242	0.0016	0.042	0.0002	0.0008

Table 5. Summary of the HAP, acetic acid, and ethanol emissions.



Figure 4. HAP emissions as a function of time.



Figure 5. HAP emissions as a function of moisture content.

The detection limits for the GC instrument were

Methanol – 1.54 µg/mL in the aqueous phase Phenol – 0.89 µg/mL in the aqueous phase Ethanol – 0.98 µg/mL in the aqueous phase Acetic acid – 4.85 µg/mL in the aqueous phase Formaldehyde - 0.05 µg/mL in the hexane phase Acetaldehyde – 0.05 µg/mL in the hexane phase Propionaldehyde – 0.05 µg/mL in the hexane phase Acrolein – 0.06 µg/mL in the hexane phase

The method detection limit varies with gas flow through the impingers, time of collection, and the amount of solution in the impingers. Based on the flow conditions and impinger volumes for 26 impinger samples, the method detection limits in the sampled gas (wet kiln exhaust) were

Methanol - 0.77-1.86 ppm Phenol - 0.15-0.37 ppm Ethanol - 0.34-0.83 ppm Acetic acid - 1.29-3.13 ppm Formaldehyde - 0.01-0.03 ppm Acetaldehyde - 0.01 - 0.02 ppm Propionaldehyde - 0.01-0.02 ppm Acrolein - mean = 0.01-0.02 ppm

Ethanol was below the detection limits in samples 19, 20, and 21. Acrolien was not detected in the first two samples, but was above detection limits in all the other samples. Propionaldehyde was below the detection limits in the fires sample. All other samples were above the detection limits. The table below shows the amounts emitted if one-half the detection limit is substituted for all samples below the detection limit. The total HAPs remain unchanged at 0.13 lb/mbf.

			Unit mass	leaving kiln				
Methanol	Phenol	Ethanol	Acetic	Form-	Acet-	Propion-	A	
Wethanor	Thenor	Luanor	acid	aldehyde	aldehyde	aldehyde	Acrolein	
lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	lb/mbf	
0.084	0.000	0.023	0.242	0.0016	0.042	0.0002	0.0008	

Field spikes were run by operating two impinger trains simultaneously. An aliquot of the compounds was added to one impinger train. Spike recovery percentage is the mass of a compound detected in the lab compared to mass added to the impinger. Table 6 shows the field spike recoveries. The method requires between 70% and 130% recovery if the concentration of the compound in the gas phase is greater than 1.5 ppm in the dry gas. The spike levels (the amount in the spiked impinger should be between three (3x) to five (5x) times that in the unspiked impinger) were under 5x

for every compound on at least one spike except for ethanol. We were attempting the "bracket method" with a high and low spike; however the concentrations of ethanol in the catch were lower than anticipated. All spike recoveries were within +/- 30% except for acetic acid (66%) on spike 9B. This acetic acid in this run was underspiked (the spike was only 0.3x the impinger catch).

The results for a field blank are shown in Table 7. None of the target compounds were detected in the blank above the detection limits. Formaldehyde was detected at approximately 60% of the detection limit and 40x lower than the typical sample concentration. Acetic acid was detected at approximately half of the detection limit and 50x lower than the typical sample concentration.

Duplicate samples were run by operating two impinger trains simultaneously. The results of duplicates are shown in Table 8. The percentage is the difference between the gas concentrations detected by each impinger. Phenol was not detected so duplicates could not be compared. Differences ranged from 2 to 30%, all within the limits of the method (+/-30%).

				Alcohol Spi					
Run	Methanol	Mass in Phenol	Ethanol	Acetic	Impinger flow	M Methanol		Ethanol	Acetic
5	µg 917.0	<u>ру</u> 0.0	уд 890.0	<u>µg</u> 5111.2	mL/min 402.5	<u>µg</u> 454.3	94 0.0	440.9	µg 2532.0
503	1039.3	142.2	5019.9	5508.3	199.4	1039.3	142.2	5019.9	5508.3
Spike			centrations		1.1.1.1.1.1	-		coveries	Acatio
mass 9	Methanol µg/mL	Phenol µg/mL	Ethanol ug/mL	Acetic µg/mL		Methanol %	Phenol %	Ethanol %	Acetic %
1	568.4	145.2	4957.7	3346.9		102.9	97.9	92.4	88.9
	577.0	149.8	4873.7	3229.0					
		P	Vdehyde S	pike	_	_			
10000	Form-	Mass in Acet-	Propion-		Impinger	Form-	Acet-	Propion-	
Run	aldehyde	aldehyde	aldehyde	Acrolein	flow	aldehyde	aldehyde	aldehyde	Acrolein
5	14.6	1606.3	2.8	14.0	mL/min 402.5	16.0	µg 1757.3	<u>µg</u> 3.1	15.4
502	31.0	2417.4	12.1	30.0	440.3	31.0	2417.4	12.1	30.0
pike mass	Form-	Spike con Acet-	Propion-			Form-		coveries Propion-	r
pike mass	aldehyde		aldehyde	Acrolein	5-0	aldehyde	aldehyde	aldehyde	Acrolain
g 0.64	µg/mL 20.4	µg/mL 842.1	µg/mL 12.9	µg/mL 20.1		% 115.0	%	% 107.9	% 113.8
				Alcohol Spi	ke				
Run	Methanol	Mass in Phenol	Ethanol	Acetic	Impinger flow	Methanol		Ethanol	Acetic
	PR	рų	Pg	Pg.	mL/min	PB	P9	Pg	PA
9 902	1820.4 3112.6	0.0 246.9	665.1 8560.2	8207.3 12976.6	400.0 439.9	2001.8 3112.6	0.0 246.9	731.4 8560.2	9025.5 12976.6
Spike	T	Spike con	centrations		1.1.1.1	1	Spike re	coveries	
mass	Methanol	Phenol	Ethanol	Acetic	Stat.	Methanol %	Phenol %	Ethanol %	Acetic %
g Senar Galass	µg/mL	µg/mL	µg/mL	pg/mL	States.	STALENY	1203621	With the lot	and the second second
1.69	779.5	144.9	5223.0	3528.9	100000	84.3	100.9	88.7	66.3
			Materia	-	i			8	
		Mass in	Aldehyde S impinger	ріке	Impinger		ass corre	cted for fk	w
Run	Form- aldehyde	Acet- aldehyde	Propion- aldehyde	Acrolein	flow	Pome	Acet- aldehyde	Propion- aldehyda	Acroleir
	Pg	PA	Pha Pha	<u>eu</u>	mL/min	Pg	Pa	Pg	P.
9 903	39.6 144.8	960.1 1947.3	4.6 25.9	24.3 11.5	400.0 199.2	19.7 144.8	478.0 1947.3	2.3 25.9	12.1 11.5
	1	Spike con	centrations	-	i antigenete	1	Spike re	coveries	
ipike mass	Form-	Acet-	Propion-	Acrolein	Cale No.	Form-	Acet-	Propion-	Acrolein
<u>9</u> 1.74	aldehyde µg/mL	aldehyde µg/mL	ug/mL	µg/mL		aldehyde %	aldehyde %	aldehyde %	%
1.74	73.0	816.7	13.4	0.0	2.	98.5	103.4	101.2	#DIV/0
	1	Mass in	impinger	Alcohol Spi	ike Impinger	1	ass corre	cted for fk	
Run	Methanol	Phenol	Ethanol	Acetic	flow	Methanol	Phanol	Ethanol	Acetic
Run 13	49 2696.1	Phenol µg 0.0	Ethanol µg 338.3	µg 6541.9	flow mL/min 412.9	Methanol µg 1330.7	Phenol µg 0.0	Ethanol µg 167.0	Acetic µg 3228.7
	49	Phenol Vg	Ethanol	рg	flow mL/min	Methanol	Phenol	Ethanol	Acetic
13 1303 Spike	µ9 2696.1 1852.9	Phenol µg 0.0 111.4 Spike con	Ethanol µg 338.3 3986.6 centrations	μg 6541.9 6258.1	flow mL/min 412.9	Methanol µg 1330.7 1852.9	Phenol µg 0.0 111.4 Spike re	Ethanol µg 167.0 3986.6 coveries	Acetic µg 3228.7 6258.1
13 1303	49 2696.1	Phenol µg 0.0 111.4	Ethanol µg 338.3 3986.6	μg 6541.9 6258.1	flow mL/min 412.9	Methanol µg 1330.7	Phenol µg 0.0 111.4 Spike re	Ethanol µ9 167.0 3986.6	Acetic µg 3228.7
13 1303 Spike mass 9	уя 2696.1 1852.9 Methanol уд/mL	Phenol µg 0.0 111.4 Spike con Phenol µg/mL	Ethanol µg 338.3 3986.6 Ethanol µg/mL	μg 6541.9 6258.1 Acetic μg/mL	flow mL/min 412.9	Methanol µ9 1330.7 1852.9 Methanol %	Phenol µg 0.0 111.4 Spike re Phenol %	Ethanol µg 167.0 3986.6 ecoveries Ethanol %	Acetic
13 1303 Spike mass	1852.9	Phenol µg 0.0 111.4 Spike con Phenol	Ethanol µg 338.3 3986.6 centrations Ethanol	µg 6541.9 6258.1 Асетіс	flow mL/min 412.9	Methanol µg 1330.7 1852.9 Methanol	Phenol µg 0.0 111.4 Spike re Phenol	Ethanol µ9 167.0 3986.6 coveries Ethanol	Acetic µg 3228.7 6258.1 Acetic
13 1303 Spike mass 9	уя 2696.1 1852.9 Methanol уд/mL	Phenol µg 0.0 111.4 Spike con Phenol µg/mL 149.8	Ethanol µg 338.3 3986.6 Ethanol µg/mL 4873.7 Ndehyde S	μg 6541.9 6258.1 Acetic μg/mL 3229.0	flow mL/min 412.9	Methanol µg 1330.7 1852.9 Methanol % 111.7	Phenol µg 0.0 111.4 Spike re Phenol % 91.9	Ethanol µg 167.0 3986.6 Ethanol % 96.8	Acetic µ9 3228.7 6258.1 Acetic % 115.8
13 1303 Spike mass g 0.81	ия 2696.1 1852.9 Метhanol µg/mL 577.0	Phenol µg 0.0 111.4 Spike con Phenol µg/mL 149.8 Mass in	Ethanol µg 338.3 3986.6 centrations Ethanol µg/mL 4873.7 Addehyde S impinger	<u>µ</u> 9 6541.9 6258.1 <u>Асеtіс</u> <u>µg/mL</u> 3229.0	flow mL/min 412.9 203.8	Methanol yg 1330.7 1852.9 Methanol % 111.7	Phenol µg 0.0 111.4 Spike re Phenol % 91.9 ass corre	Ethanol µg 167.0 3986.6 ecoveries Ethanol %	Acetic µg 3228.7 6258.1 Acetic % 115.8 ww
13 1303 Spike mass 9	US 2696.1 1852.9 Methanol yg/mL 577.0	Phenol yg 0.0 111.4 Spike con Phenol yg/mL 149.8 / Mass in Acet- aldehyde	Ethanol H3 338.3 3986.6 centrations Ethanol µg/mL 4873.7 Vdehyde S impinger Propion- aldehyde	<u>µg</u> 6541.9 6258.1 Асеtіс <u>µg/mL</u> 3229.0 ріке Астоlein	flow mL/min 412.9 203.8	Methanol µg 1330.7 1852.9 Methanol % 111.7 N Form- aldehyde	Phenol µg 0.0 111.4 Spike re Phenol % 91.9 ass corre Acet- aldehyde	Ethenol J9 167.0 3986.6 Ethanol % 96.8 ethed for fix Propion- aldehyde	Acetic µg 3228.7 6258.1 Acetic % 115.8 ww Acrolein
13 1303 Spike mass g 0.81 Run 13	ия 2696.1 1852.9 Метлапої µg/mL 577.0 577.0	Phenol yg 0.0 111.4 Spike con Phenol yg/mL 149.8 Mass in Acet- aldehyde yg 776.7	Ethanol <u>49</u> 3383.3 3986.6 centrations Ethanol <u>4873.7</u> Addenyde S impinger Propion- aldenyde <u>49</u> 4.9	µg 6541.9 6258.1 Асеtіс µg/mL 3229.0 ріке Асгоlеіл µg 20.1	flow mU/min 412.9 203.8 Impinge/ flow mU/min 412.9	Меthanol	Phenol µg 0.0 111.4 Spike re Phenol 96 91.9 91.9 Pass corre Acet- akdehyde µg 843.6	Ethenol yg 167.0 3996.6 coveries Ethanol % 96.8 cted for fik Propion- akdehyde yg 5.3	Acetic <u>J</u> 9 3228.7 6258.1 Acetic % 115.8 ow Acrolein <u>J</u> 9 21.8
13 1303 Spike mass g 0.81 Run	<u>у</u> 9 2696.1 1852.9 Меthanol уд/mL 577.0	Phenol PB 0.0 111.4 Spike con Phenol µg/mL 149.8 Ass in Acet- aldehyde µg	Ethanol µ9 338.3 3986.6 centrations Ethanol µg/mL 4873.7 Ndehyde S impinger Propion- aldehyde µ9	<u>µg</u> 6541.9 6258.1 Асетіс <u>µg/mL</u> 3229.0 ріке Астоlein µg	flow mL/min 412.9 203.8 Impinger flow mL/min	Methanol yg 1330.7 1852.9 Methanol % 111.7 N Form- aldehyde yg	Phenol µg 0.0 111.4 Spike m Phenol % 91.9 ass corre Acet- aldehyde µg	Ethanol J9 157.0 3986.6 Ecoveries Ethanol % 96.8 Propion- aldehyde J9	Acetic µg 3228.7 6258.1 Acetic % 115.8 ww Acrolein µg
13 1303 Spike mass 9 0.81 Run 13 1302	ия 2696.1 1852.9 Меthanol уд/mL 577.0 Form- aldehyde ия 48.9 102.0	Phenol µg 0.0 111.4 Spike con Phenol µg/mL 149.8 Mass in Acet- aldehyde µg 776.7 1517.6 Spike con	Ethenol H9 338.3 3986.6 Ethanol µg/mL 4873.7 Vdehyde S impinger Propion- aldehyde 49 14.3 centrations	µg 6541.9 6258.1 Асеtic µg/mL 3229.0 ріке Асгоlеіл µg 20.1 23.0	flow mU/min 412.9 203.8 Impinge/ flow mU/min 412.9	Methanol µg 1330.7 1852.9 Methanol % 111.7 Methanol % 111.7 Methanol % 111.7 Methanol % 111.7	Phenol yg 0.0 111.4 Spike m Phenol % 91.9 91.9 stars corre Acet- akteryde yg 843.6 1517.6 Spike m	Ethanol yg 167.0 3986.6 Ethanol % 96.8 eted for fk Propion- akdehyde yg 5.3 14.3 ecoveries	Acetic <u>J</u> 9 3228.7 6258.1 Acetic % 115.8 ow Acrolein <u>J</u> 9 21.8
13 1303 Spike mass P 0.81 Run 13 1302 Spike mass	ид 2696.1 1852.9 Меthanol µg/mL 577.0 Form- aldehyde 102.0	Phenol Phenol Phenol Phenol pg/mL 149.8 Mass in Acet- aldehyde Phenol pg/mL 149.8 Mass in Acet- aldehyde Acet- aldehyde	Ethanol H9 338.3 3986.6 Ethanol H2 4873.7 Vdehyde S implinger Propion- aldehyde H9 14.3 Centrations Propion- aldehyde	μg 6641.9 6258.1 Acetic μg/mL 3229.0 3200.0 3229.0 32000	flow mU/min 412.9 203.8 Impinge/ flow mU/min 412.9	Меthanol	Phenol yg 0.0 111.4 Spike rr Phenol % 91.9 91.9 91.9 843.6 1517.6 Spike rr Acet- aldehyde	Ethanol UQ 157.0 3986.6 Ethanol 5% 96.8 96.8 Propion- aldehyde UQ 5.3 14.3 propion- aldehyde	Acetic <u>J</u> 9 3228.7 6258.1 Acetic % 115.8 ow Acrolein <u>J</u> 9 21.8
13 1303 Spike mass 9 0.81 Run 13 1302 Spike mass 8	V3 2696.1 1852.9 Methanol yg/mL 577.0 577.0 577.0 Form- aldetyde V3 46.9 102.0	Phenol µg 0.0 1111.4 Spike con Phenol µg/mL 149.8 Acet- aldehyde µg 776.7 1517.6 Spike con Acet- aldehyde µg/mL	Ethenol µg 338.3 3986.6 Centrations Ethanol µg/mL 4873.7 Mdehyde S impinger Propion- aldehyde µg 14.3 Centrations Centrations Propion- aldehyde µg/mL	µg 6541.9 6258.1 Асеtic µg/mL 3229.0 ріке Асгоlеіл µg 20.1 23.0	flow mU/min 412.9 203.8 Impinge/ flow mU/min 412.9	Methanol µg µg 1330.7 1330.7 1852.9 Methanol % 111.7 111.7 Form-addehyde µg 53.1 102.0 Form- Form-	Phenol Hg 0.0 111.4 Spike rr Phenol % 91.9 Phenol % 91.9 Acet- aldehyde Hg 843.6 1517.6 Spike rr Acet- aldehyde %	Ethanol H9 167.0 3986.6 Ethanol % 96.8 Propion- aldehyde H9 5.3 14.3 scoveries Propion- aldehyde %	Acetic
13 1303 Spike mass P 0.81 Run 13 1302 Spike mass	ид 2696.1 1852.9 Меthanol µg/mL 577.0 Form- aldehyde 102.0	Phenol Phenol Phenol Phenol pg/mL 149.8 Mass in Acet- aldehyde Phenol pg/mL 149.8 Mass in Acet- aldehyde Acet- aldehyde	Ethanol H9 338.3 3986.6 Ethanol H2 4873.7 Vdehyde S implinger Propion- aldehyde H9 14.3 Centrations Propion- aldehyde	µg 6541.9 6258.1 Асетіс µg/mL 3229.0 3229.0 ріке Астоlein µg 20.1 23.0 Астоlein µg/mL	flow mU/min 412.9 203.8 Impinge/ flow mU/min 412.9	Methanol µg 1330.7 1852.9 Methanol % 1111.7 N Form- aldehyde µg 53.1 102.0	Phenol yg 0.0 111.4 Spike rr Phenol % 91.9 91.9 91.9 843.6 1517.6 Spike rr Acet- aldehyde	Ethanol H9 167.0 3986.6 Ethanol % 96.8 Propion- aldehyde H9 5.3 14.3 scoveries Propion- aldehyde %	Acetic µ9 3228.7 6258.1 Acetic % 115.8 0w Acroleir µ9 21.8 23.0
13 1303 Spike mass 9 0.81 Run 13 1302 Spike mass 8	V3 2696.1 1852.9 Methanol yg/mL 577.0 577.0 577.0 Form- aldetyde V3 46.9 102.0	Phenol µg 0.0 111.4 Spike con Phenol µg/mL 149.8 // Mass in Acet- aldehyde µg 776.7 1517.6 Spike con Acet- aldehyde µg/mL 816.7	Ethanol JB 338.3 3966.6 Centrations Ethanol Lethanol 4873.7 Adenyde S impinger Propion- aldetyde J9 14.3 Centrations Propion- aldetyde J9 14.3 Centrations Propion- aldetyde J9 14.3 Centrations Centrati	µg 6541.9 6258.1 Асетіс µg/mL 3229.0 3229.0 ріке Астоlein µg 20.1 23.0 Астоlein µg/mL	flow mL/min 412.9 203.8 Impinger flow mL/min 412.9 448.5	Methanol µg 1330.7 1852.9 Methanol % 111.7 N Form- aldehyde µg 53.1 102.0 Form- aldehyde % 99.9	Phenol Hg 0.0 111.4 Spike re Phenol % 91.9 843.6 1517.6 Spike re Acet- aldehyde % 123.2	Ethanol µg 157.0 3986.6 Ethanol % 96.8 96.8 96.8 96.8 96.8 96.8 14.3 ropoin- aldehyde ¥g 5.3 14.3 Propion- aldehyde % 100.5	Acetic P9 3228.7 6258.1 Acetic % 115.8 300 Acroleir P9 21.8 23.0 Acroleir % Acroleir %
13 1303 Spike mass 9 0.81 Run 13 1302 Spike mass 8	vg 2696,1 1852.9 Methanol yg/mL 577.0 Form- aldehyde yg 46.9 102.0 Form- aldehyde yg 73.0	Phenol µg 0.0 111.4 Spike con Phenol µg/mL 149.8 Acet- aldehyde µg 776.7 1517.6 Spike con Acet- aldehyde µg/mL 816.7 Mass in Acet- aldehyde µg/mL	Ethanol H3 338.3 3966.6 centrations Ethanol Hg/mL 4873.7 Vdehyde S impinger Propion- aldehyde H3 A.9 14.3 centrations Propion- aldehyde H3 Centrations Propion- aldehyde H3 Centrations Propion- aldehyde H3 Centrations Centrat	<u>µ</u> 9 6541.9 6258.1 Асетіс <u>µ</u> 9/mL 3229.0 Ріке Асгоівіл <u>µ</u> 9 20.1 23.0 Асгоівіл <u>µ</u> 9 Асгоівіл <u>µ</u> 9	flow mL/min 412.9 203.8 Impinger flow mL/min 412.9 448.5	Methanol µg 1330,7 1852,9 Methanol % 111,7 111,7 N Form- aldehyde µg 53,1 102,0 Form- aldehyde 99,9 M	Phenol Mg 0.0 111.4 Spike m Phenol % 91.9 91.9 ass corre Acet- aldehyde M3.6 843.6 843.6 843.6 843.6 843.6 Spike m Acet- aldehyde % 1517.6 Spike m Acet- aldehyde % 123.2	Ethanol J93 157.0 3986.6 Ethanol 54 96.8 Propion- aktehyde J93 14.3 scoveries Propion- aktehyde J9 14.3 scoveries Scoveries Propion- aktehyde Scoveries Sco	Acetic P9 3228.7 Acetic % 115.8 115.8 200 Acroleir 90 21.8 23.0 Acroleir % #DIV/00 50 50 50 50 50 50 50 50 50
13 1303 Spike mass g 0.81 Run 13 1302 Spike mass g 0.67 Run	93 2696.1 1852.9 Methanol yg/mL 577.0 Form-aldetryde 99 48.9 102.0 Form-aldetryde 99/mL 73.0 Methanol	Phenol yg 0.0 111.4 Spike con Phenol yg/ml, 149.8 // Mass in Acot- aldehyde yg/ml, 816.7 Mass in Phenol yg/ml, 816.7	Ethanol Hg 338.3 3986.6 certrations Ethanol Hg/mL 4873.7 Ndehyde S implinger Propon- aldehyde Hg 4.9 14.3 certrations Propon- aldehyde Hg/mL 13.4 implinger Ethanol Hg		flow mUmin 412.9 203.8 Impinger flow ML/min 448.5	Меthanol µg 1330,7 1852,9 Меthanol % 111,7 Голт- aldehyde µg 53,1 102,0 Голт- aldehyde % 99,9 Меthanol µg	Phenol 9 Jug 0.0 111.4 Spike re and the second s	Ethanol yg 157.0 3996.6 coveries Ethanol 94. 96.8 eted for fit Propion- aldehyde yg 5.3 14.3 coveries Propion- aldehyde % 100.5	Acetic yg 3228.7 6258.1 Acetic % 1115.8 23.0 Acroleir yD X.8 23.0 Acroleir % yD V/0 % X.8 Acetic % % Acroleir % % % % % % % % % % % % %
13 1303 Spike mass 9 0.81 Run 13 1302 Spike mass 9 0.67	vg 2696.1 1852.9 Methanol yg/mL 577.0 Form- aldetryde yg/mL 577.0 Form- aldetryde yg/mL 73.0 Methanol yg/mL 2137.6	Phenol 90 90 00 00 00 00 00 00 00 00 00 00 00 00 0	Ethenol yg 338.3 3386.6 certratione Ethenol µg/mL 4873.7 Ndehyde S impinger Propion- aldehyde µg 14.3 certratione Propion- aldehyde µg 14.3 certratione Propion- aldehyde µg 14.3 certratione Propion- aldehyde µg 70.0 µg 70.0 Propion- aldehyde µg 70.0 µ	нд 6541.9 6258.1 6258.1 Acetic μg/mL 3229.0 Pike Acrolein μg 20.1 23.0 Acrolein μg 20.1 Acrolein μg 0.0 Acotol Spi Acetic μg 3980.2	flow mU/min 412.9 203.8 Impinger flow mU/min 412.9 448.5 Ke Impinger flow mU/min flow	Methanol Methanol µg 1330,7 1852.9 Methanol Methanol 5 111,7 N Form-aldehyde 9 J331,102.0 Form-aldehyde Form-aldehyde 53,1 J02.0 Form-aldehyde Methanol 99,9 J10250 N	Phenol yg 0.0 0.111.4 Spike nr 91.9 91.9 91.9 91.9 93.9 93.9 94.36 1517.6 Spike nr Acat- aldehyde 1517.6 Spike nr Phenol 91.9 93.9 94.9 94.9 94.9 95.9	Ethanol µg 167.0 3966.6 coveries Ethanol \$ 96.8 96.8 coveries Ethanol \$ 96.8 coveries \$ 96.8 coveries \$ 96.8 \$ 96.8 \$ 96.8 \$ 96.8 \$ 96.8 \$ 96.8 \$ 96.8 \$ 96.9 \$ 14.3 \$ \$ \$ \$ \$ \$ \$ \$ \$ \$ \$ \$ \$	Acetic 49 3228.7 6258.1 Acetic 56 115.8 21.8 21.8 23.0 Acroleir 92 21.8 23.0 Acroleir 95 21.8 23.0 Acroleir 96 21.8 20.0 7 4 20.0 2
13 1303 Spike mass 9 0.81 Nun 13 1302 Spike mass 9 0.67 Run 17 1703	93 2696.1 1852.9 Methanol yg/mL 577.0 Form-aldetryde 99 48.9 102.0 Form-aldetryde 99/mL 73.0 Methanol	Phenol y yg 0.0 111.4 Spike con Phenol pg/mL 149.8 776.7 Spike con 2776.7 1517.6 Spike con 276.7 1517.6 Spike con 276.7 Spike con 276.7 S	Ethanol yg 3383,5 3986,6 Contration gg/mL 4873,7 Fropon- pg/mL 4873,7 Fropon- pg/mL 14,3 Contration Propion- gg/mL 14,3 Contration Propion- gg/mL 13,4 4,9 14,3 Contration Propion- pg/mL 13,4 Contration Propion- 20,5 Contration- 20,5 Contration- 20		flow mUmin 412.9 203.8 Impinger flow ML/min 448.5	Меthanol µg 1330,7 1852,9 Меthanol % 111,7 Голт- aldehyde µg 53,1 102,0 Голт- aldehyde % 99,9 Меthanol µg	Phenol yg 00 111.4 Spike nr Phenol 5 91.9 91.9 91.9 91.9 91.9 91.9 91.9 91	Etheracial yg 187.00 source and source and source and source and source and yg source and source and so	Acetic 19 3228.7 6258.1 Acetic % Acrolein 9 21.8 23.0 Acrolein % #DIV/00 % Acetic 115.8 30 30 30 30 30 30 30 30 30 30
13 1303 Spike mass 9 0.81 Run 13 1302 spike mass 9 0.67 Run 17	93 2696.1 1852.9 Methanol pg/ml 577.0 Form- aldetyde 93 102.0 Form- Form- aldetyde 93 102.0 Form- aldetyde 93 102.0 Form- aldetyde 93 102.0 Form- Form- Barto 93 102.0 Form- Barto 93 102.0 Form- Form	Phenol U yg 0.0 111.4 Spike conn Phanol pg/mL 149.8 Mass in Acet- aldehyde pg/mL Spike conn Phanol pg/mL 149.8 Spike conn Phanol pg/mL 149.8 Spike conn Phanol pg/mL 149.8 Spike conn Phanol pg/mL 149.8 Spike conn Phanol pg/mL 151.6 Spike conn Spike conn Spike conn Spike conn Phanol pg/mL Spike conn Spike conn Phanol Phan	Ethanol 19 338.3 3986.6 Sorfrations Ethanol 19 19 19 19 19 19 19 19 19 19	ug ug 6541.9 Acetic gymL 3229.0 acrolein ug 20.1 20.1 Acrolein ug 20.1 Acrolein Acrolein acrolein	flow mU/min 412.9 203.8 Impinger flow mU/min 412.9 448.5 Ke Impinger flow mU/min flow	Metranol 99 (1330,7,7) (1852,9) (199,10	Phenol 99 99 00 111.4 Spike nr Phenol 9 56 91.9 91.9 91.9 91.9 91.9 91.9 91.9 91.	Ethanol J yg 187.00 icoveries Echanol Second 54.6 96.8	Acetic 93 92 92 92 92 92 93 228.7 6256.1 96 96 96 97 97 97 97 97 97 97 97 97 97
13 1303 Spike mass 0.81 0.81 0.81 13 1302 Spike mass 9 0.67 Run 17 1703 Spike	93 26996.1 1852.9 Methanol 577.0 577.0 Form- aldehyde 99 102.0 Form- aldehyde 92 102.0 Form- aldehyde 92 102.0	Phenol J J9 0.0 111.4 Spike con Phenol Phenol pg/mL 149.8 Aost- aldehyde J9 776.7 1517.6 Spike con Spike con Mass in Phenol J9 Mass in Spike con Spike con	Ethanol 19 3383.3 3986.6 Ethanol 1927 1978 1978 1978 1978 1978 1979 14.3 1078 14.3 11.4		flow mU/min 412.9 203.8 Impinger flow mU/min 412.9 448.5 Ke Impinger flow mU/min flow	Metanol 99 (1330 7 (1852 9 Metanol % 111.7 Porm- aldehyde 50.1 50.1 50.1 50.1 50.1 50.1 50.1 50.1	Phenol 949 949 0.0 111.4 Spike nr Phenol 95 91.9 91.9 91.9 91.9 91.9 91.9 91.9 9	Ethanol y ya 1970 - 2000 19870 6 coveries Ethanol - 2000 -	Acetic 93228.7 6256.1 Acetic 5 4 Acrolein 9 21.8 21
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Table 6. Spike test results.

Columbia Vista, Douglas-fir

	Field blank										
Mothanal	Dhanal	Ethanol	Acetic	Form-	Acet-	Propion-	A enelsis				
Methanol	Phenol	Ethanor	acid	aldehyde	aldehyde	aldehyde	Acrolein				
ppm	ppm	ppm	ppm	ppm	ppm	ppm	ppm				
0.000	0.000	0.000	2.297	0.030	0.000	0.000	0.000				

Table 7. Results for the field blank.

Table 8. Results for duplicate runs.

	1			Mass in	impinger				
Run	Methanol	Phenol	Ethanol	Acetic	Form-	Acet-	Propion-	Acrolein	Impinge flow
rtan	Wethanor	THEIR	Linarior	acid	aldehyde	aldehyde	aldehyde	ACIOIEIII	now
	рд	μg	μg	μg	рд	μg	μg	μg	mL/min
6	1054.2	0.0	733	5288.3	16.1	894.2	2.5	13.5	400.5
602	1178.0	0.0	798	5898.6	18.4	1031.1	2.9	15.4	442.7
Difference, %	1.1	#DIV/0!	1.6	0.9	3.1	4.2	4.2	3.1	

				Mass in	impinger				1
Run	Methanol	Phenol	Ethanol	Acetic	Form-	Acet-	Propion-	Acrolein	Impinger
Run	Wethanor	FILEIIUI	Ethanor	acid	aldehyde	aldehyde	aldehyde	Acrolein	flow
	рд	μg	μg	μg	μg	μg	μg	μg	mL/min
10	2148.0	0.0	628.3	7776.1	36.4	718.1	3.5	17.7	401.8
1002	2159.0	0.0	702.0	7967.9	40.1	800.6	3.9	20.1	440.2
Difference, %	8.6	#DIV/0!	2.0	6.7	0.6	1.8	1.6	3.1	

				Duplicate	9				
				Mass in	impinger				1
Run	Methanol	Phenol	Ethanol	Acetic	Form-	Acet-	Propion-	A	Impinge
Run	Wethanor	Phenoi	Ethanor	acid	aldehyde	aldehyde	aldehyde	Acrolein	flow
	hà	μg	μg	μg	μg	μg	μg	μg	mL/min
14	2579.9	0.0	208.6	5983.7	43.5	601.2	4.0	15.9	409.4
1402	3112.7	0.0	167.5	7692.8	55.1	777.5	5.0	20.0	444.7
Difference, %	10.5	#DIV/0!	30.0	16.8	15.5	17.4	13.9	14.7	

4. Control system and operating conditions

A schematic of the kiln is shown in Figure 6 (top). The kiln box is approximately 4' by 4' by 4'. It is indirectly heated by steam. Four dry-bulb thermocouples and two wet-bulb thermocouples are located on the entering-air side of the load. The dry-bulb thermocouples are spaced in a grid. The two wet-bulb thermocouples are under a single sock at the center of the entering-air side of the load.

Humidity control

A 200 L/min MKS mass flow meter controlled the amount of air entering the kiln. It was factory calibrated and checked using a bubble meter. The amount of air entering the kiln is based on the wet-bulb temperature - if it is above setpoint, the airflow is increased and if it is below setpoint the airflow is decreased. This is analogous to venting for a commercial kiln. A minimum of 2 L/min entered the kiln at all times, more than removed by the analyzer (1.6 L/min). Putting air into the kiln at a rate of 100 L/min causes the pressure in the kiln to be 60 to 130 Pa above ambient, depending on location in the kiln (high-pressure or low-pressure side). Thus, any fugitive leakage should be out of the kiln. Two additional flow meters can be manually set to provide additional airflow.

Temperature control

Temperature in the kiln is controlled by indirect steam heating. When the dry-bulb temperature is below setpoint, the steam pressure in the coil is increased. When it is above setpoint, steam flow to the coil is reduced.

The dry- and wet-bulb temperatures recorded for each charge are shown in Figure 7. The schedule provided by the mill is also shown. The disturbance at 108 hours is when the kiln was opened for a moisture content check. The kiln was unable to maintain wet-bulb temperature towards the end of the schedule due to the slow rate of water release from the boards.





Figure 6. Schematic of kiln and sampling system (top) and photos of charge in kiln (bottom).

5. Production-related parameters

Wood quantity

The wood properties were determined using the nominal wood dimensions (16/4 in this case) which provides for 1.66 board feet per lineal foot of board. There were 21 pieces in the kiln at 44" in length. The sum of the 42 boards was 77 feet. The board footage was therefore 127.8 board feet. This quantity was used to express the emissions from the drying cycle on a production basis of lb/mbf (pounds per thousand board feet).

Wood properties

The wood property measurements are shown in Table 9. Individual measurements can be found in the Excel file "Weights, CV-1.XLS" in Appendix 2.

There was no included pith due to the lumber grade. There was only one board with visible pitch pockets.

The initial moisture content (39%) was close to that published for Douglas-fir (37%) in the USDA Wood Handbook.

The average ring count was determined by counting the rings over a 1" radial distance and averaging for all boards.

The number of knots were counted on the top face of each board and averaged. Knot diameter is an average of the knots present. The knots occupied approximately 0.1% of the boards' faces.

Table 9. Wood properties.

Kn	ots	Heartwood	Ring	Pith
Number	Diameter		Count	In
#	in	%	#/inch	Y/N
1.2	0.5	92	6.3	0



Figure 7. Schedules followed (top) and schedule provided by mill (bottom).

6. Test methods

Charge Sequence

The lumber was unwrapped and 2" were trimmed from each end of each board to give 44" samples. These were then weighed, placed in the kiln and dried. At the end of drying the wood was weighed, oven dried, and reweighed so initial and final moisture contents could be determined by ASTM D4442 (oven-dry method).

Sampling Methodologies

Hydrocarbon

Sampling for total hydrocarbon is done directly from the kiln as shown in Figure 6 (top). The concentration obtained from the hydrocarbon analyzer and the amount of air entering the kiln allow the total hydrocarbon emissions to be calculated.

Figure 8 shows the hydrocarbon sampling system. Unlike stack testing, all necessary equipment is located in a lab and flows are controlled with valves. The sample is withdrawn from the kiln under the assumption that the gas in the kiln is well-mixed and that the composition in the kiln near the exhaust is the same as the composition of the exhaust. The THC sample was drawn from the kiln into a heated dilution/filter box. The box was heated to 235-245°F. Heated dilution gas was added to the hydrocarbon sample gas to lower the gas moisture content to the detector (except for runs 1 and 2) so that the air moisture content to the detector remained less than 15%. The sample line from the box to the analyzer was heated to 245°F.

The fuel gas was hydrogen. The span gas was EPA Protocol 197 ppm propane in air, the mid-gas was EPA Protocol 50 ppm propane in air. The zero gas was <0.1 ppm air. Detailed sampling procedures are in Appendix 1.

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Figure 8. Schematic of heated filter box with air dilution system, heated sample line, and analyzer (top). Sample enters heated box from back of drawing through a heated sampling line. Calibration gas valves and dilution air valves are to left. Line to analyzer is the green line on the left in the left photo.

HAPs

The sampling train for NCASI Method 105 is shown in Figure 9. The impingers were in a glycol solution maintained at -1 C. Prior to each sampling interval, the impingers were laboratory-washed and 10 to 15 mL of BHA solution were added

to the first and second impingers. The third impinger was left empty. The fourth impinger was present in the system to prevent any overflow from reaching the critical orifice. The system was then assembled and a vacuum check was performed with the valves at each end closed. Less than 1" Hg of pressure change over 2 minutes was acceptable. This was met for each interval. The flow rate through the system was then measured using a Gilibrator flow meter to take four flow readings at the probe tip. This was approximately 200-500 mL/min, depending on the sampling train. A valve at the probe tip was then turned to sample from the kiln and the sampling interval begun. The collection interval time was approximately 1:30 and an interval was started approximately every six hours.

The flow rate was measured after each sampling interval. The fluid in the three impingers was weighed and placed in a glass bottle. The impingers were then rinsed with 10 mL of water followed by 3 to 5 mL of hexane. The rinses were also placed in the bottle and it was sealed. Samples were kept refrigerated and in the dark until lab analysis was done. Lab analysis was done within one week of sample collection.

The local airport altimeter setting and the lab temperature were recorded at the beginning and end of each interval so the flow rates could be adjusted to standard conditions.





7. Analytical procedures

Hydrocarbon

Leak checks of the VOC sampling train were conducted before and after the charge was dried. A valve was closed at the probe tip and a 3-way valve was closed at the back of the analyzer. All components from just behind the probe tip to the valve at the back of the analyzer were placed under a 15-20 inHg vacuum. Less than one inHg pressure change during two minutes is acceptable and this was met.

Total flow and sample flow to the analyzer were checked using an NIST-traceable flow meter. Total flow is measured with the dilution gas off and is equal to both the sample flow from the kiln when the dilution is off and the total volume drawn by the analyzer. Sample flow is measured with dilution gas on (if used for that interval) and is the volume of gas sampled from the kiln when the dilution gas is on. This was done at the beginning and end of each sampling interval. The meter was attached to the system near the probe tip within the heated box. The valves were repositioned so that the sample came from the flow meter rather than the kiln. Readings of flow were made with the dilution gas both off and on. The flow readings were verified by observing the analyzer reading for span gas with the dilution gas off and on. The dilution was not actually used for runs 1 and 2 because the kiln wet-bulb was low enough initially that the gas moisture content was less than 15% until the wet-bulb temperature was greater than 130-135°F.

Calibration of the zero and span of the detector was done at the beginning of each run (about every six hours). The calibration gas was introduced by setting the valves so the calibration gas entered the system in the white heated mixing box at ambient pressure. The calibration was checked at the end of each run with no adjustments made to the instrument's zero or span during the run. A span drift less than 10% of the span value was acceptable. A zero drift of less than 3% of the span value was acceptable. A total calibration drift less than 10% was acceptable for a sampling run. These criteria were met.

HAPs

Lab analysis for aldehydes

Aldehyde standards were prepared by the dilution of neat aldehydes in water (to 400 ppm for formaldehyde, propionaldehyde, acrolein and acetaldehyde). This stock solution was mixed a BHA solution [from ortho-benzylhydroxylamine hydrochloride (BHA) and deionized water (30g BHA per liter of water)]. The stock solution-BHA mixture was vigorously agitated and allowed to sit for six hours to allow for derivatization of the aldehydes into aldoximes. The derivatized aldehyde solution was extracted with three aliquots of hexane to create a 300 ppm stock solution in hexane. This was diluted by mass to make standards down to 0.25 ppm. 1.8 mL aliquots were place in GC autosampler vials with 10 μ L of 27,830 ppm nitrobenzene added to each as an internal standard.

The samples (from the bottles collected in field) were prepared by performing three extractions in a separatory funnel. The first extraction was with the hexane added in the field plus a 3.5 mL aliquot. The second extraction was with a 7-mL aliquot of hexane again placed in the jar with the liquid fraction. The final extraction was done with 7 mL of clean hexane in the separatory funnel after rinsing the jar with the hexane. The total hexane volume was approximately 20 mL. The volumes of the two phases were calculated from their weights. A 1.8 mL aliquot of the hexane fraction was transferred to an autosampler vial and spiked with internal standard.

The analytical instrument was a Shimadzu GC model 2010 with a flame thermionic detector (FTD), the Shimadzu equivalent of a nitrogen phosphorous detector (NPD). The column was a 105-meter Restek RTX-5 capillary with a 0.25 mm outside diameter and a stationary phase thickness of 0.25 μ m. The oven schedule was: 2 minutes at 120°C, 2°C/min ramp to 160°C, 40°C/min ramp to 220°C and 6.5 minutes at 220°C. The column flow was 25 cm/sec, with 3 mL/min septum purge, and a 1:10 split ratio with a glass wool packed split injection liner. The detector make up He was set to 20 mL/min and the H₂ was set to 3 mL/min. The air was set to 140 mL/min, and the source current was set to 2 pA. The He and H₂ gases were grade 5 and the air was grade 0.1. The injector temperature was 200°C and the detector temperature 280°C. An AOC-20i autosampler was used to perform 1 μ L injections using a 10 μ L syringe with a steel plunger.

Lab analysis for alcohols

Standards for methanol, phenol, ethanol, and acetic acid were prepared by the volumetric dilution of neat reagents in water. The mixed standard was prepared at a concentration of 900 milligrams per liter (mg/L). Additional standards were prepared by the volumetric dilution of the mixed standard at a range from 0.5 mg/L to 500 mg/L. Aliquots of these were placed into autosampler vials with 10 μ L of 29,455 ppm cyclohexanol internal standard.

Samples were prepared by transferring aliquots of the previously hexane-extracted aqueous fractions into autosampler vials and adding internal standard. The analytical instrument was a Shimadzu GC model 2010 with a FID detector. The column was a 60-meter Restek Stabilwax capillary with a 0.53 mm outside diameter and a stationary phase thickness of 1.5 μ m. The oven schedule was: 3 minutes at 60°C, 10°C/min ramp to 80°C, 3 minutes at 80°C, 10°C/min ramp to 230°C, and 10 minutes at 230°C. The column flow was 30 cm/sec, with 3 mL/min septum purge, and a 1:10 split ratio with a glass wool packed split injection liner. The detector make up He was set to 25 mL/min and the H₂ was set to 50 mL/min.

The air was set to 500 mL/min. The He and H₂ gases were grade 5 and the air was grade 0.1. The injector temperature was 175°C and the detector temperature 250°C. An AOC-20i autosampler was used to perform 1 μ L injections using a 10 μ L syringe with a PTFE plunger.

8. Field data sheets and sample calculations

Field data sheets

Samples of field data sheets are shown in Figures 10 to 13. All field data sheets are in Appendix 2 this report in electronic format (pdf).

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Figure 10. Sample of field data sheet for hydrocarbon analyzer.

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Figure 11. Sample of field data sheet for HAPs collection.

Columbia Vista, Douglas-fir

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						End	<u> </u>											
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1:54	14:59	11	175	152	750	76=	= 39	-1	1.38	-	-	1.4	-	-	-	-	-	

Figure 12. Sample of kiln log data sheet.



Figure 13. Sample of flow measurement record.

Calculations

The "FlowCalc" worksheet in the Excel files "Kiln, CV-1.XLS" in Appendix 2 shows the calculations for each 3-minute interval during the charges. Column A is a reading number. Columns B and C are the clock and charge times, respectively. Columns D/E and F/G are the average dry- and wet-bulb temperatures.

Humidity

Column H is the vapor pressure (P_{vp} , Pa) of water at the wet-bulb temperature. The absolute humidity (AbHum, kg_{water}•kg_{air}-1) is shown in column I and the molal humidity (mol_{water}•mol_{air}-1) in column J. These are calculated based on the dry-bulb temperature (T_d, °C) and wet-bulb temperature (T_w °C),

 P_{vp} , = $P_{ambient}$ * 10^{(16.373 - 2818.6/(Td+273.16)} - 1.6908*LOG10(Td+273.16) - 1.6908*LOG10(Td+273.16) - 1.6908*LOG10(Td+273.16)

0.0057546*(Td +273.16) + 0.0000040073*(Td +273.16)**2)

AbHum = (MW_{water} / MW_{air}) * (1 / (P_{kiln}/P_{vp}-1)) - ((T_d-T_w) * R_{psy}) / λ

MolHum = AbHum * MWair / MWwater

where MW are molecular weights (kg•kgmol⁻¹), R_{psy} is the psychrometric ratio (0.95 kJ•kg⁻¹•K⁻¹), and λ is the latent heat (2419 kJ•kg⁻¹).

Flows

The volumetric dry gas flow rate (DryGasV, L•min⁻¹) in column K is the flowmeter reading adjusted for the meter calibrations and the molar humidity of the entering gas. This is in standard (at 0°C) liters per minute. In column L this has been converted to a mass flow rate (DryGasM, kg•min⁻¹) and in column M is the same information is expressed as a molal flow rate (DryGas, kgmol•min⁻¹). These values are for the dry gas vented from the kiln.

DryGasV = (FlowMeter1 + FlowMeter2 + FlowMeter3) * (1/(1+MolHumin))

 $DryGasM = (DryGasV L \cdot min^{-1}) * 1/(22.4 m^{3} \cdot kgmol^{-1}) * MWair / (1000 L \cdot m^{-3})$

DryGas (kgmol/min) = DryGasM / MWair

The water removal rate (WaterVented, g•min⁻¹) (column N) is calculated from the humidity (column I) and the gas flow (column L). The total water (column O) is an integration of column N over time.

WaterVented = (AbHum - AbHum_{in}) * (DryGasM * 1000 g•kg⁻¹)

Moisture content

The moisture content of the wood at each three-minute interval (column P) was determined by reducing the moisture content of the wood from the previous value by accounting for the amount of water leaving the kiln during the interval.

 $MC = MC_{Previous} - 100 * (WaterVented / (1000 g \cdot kg^{-1}) / ODWoodWt)$

This amount is then adjusted by adjusting the wet-bulb temperature to make the ending moisture content match that measured by ASTM D4222.

Hydrocarbon

The original total hydrocarbon analyzer reading is shown in column Q. In column R this has been corrected to compensate for the range setting switch on the analyzer. Also in column R, the THA data between sampling runs (rows labeled "test" in column AA) has been adjusted to the average of the data during the 9-minute period before and the 9-minute period after the analyzer testing and calibration time.

The dilution THA (column S) is the corrected THA reading divided by the dilution ratio (from column AA). In column T we have the opportunity to compensate for the effect of moisture on the JUM detector. Column T equals column S because dilution was used and no compensation was made. Finally in column U, the hydrocarbon concentration is converted to a dry gas basis concentration using the molar humidity (column J).

THC_{Dry}, ppm= THC * (1 + MolHum)

In column V, the hydrocarbon flow rate (THC_{Vented}, g_{Carbon}•min⁻¹) is calculated in a manner analogous to the water flow rate using the dry gas flow rate and the hydrocarbon concentration.

THCvented = DryGas * (THC_{Dry} / 10⁶) * MW_{Propane} * (1000 g•kg⁻¹) * (0.81818 g_{Carbon}•g_{Propane}⁻¹)

Column W is the integral of column V over time, the cumulative hydrocarbon released up to that point in the schedule (in grams). Column X is the cumulative unit emissions, that is, column W divided by the oven-dry weight of the wood in the kiln. Column AI is the cumulative emissions in pounds per thousand board feet and column AH is the rate of emissions release (lb•mbf⁻¹•hr⁻¹)

Column Z indicates the hydrocarbon sampling run and column AA is the dilution ratio during that run.

The remaining columns are used not used in the hydrocarbon calculations. They are for graphing shown on other worksheets in the workbook.

At the end of the FlowCalc spreadsheet (at the bottom) are summaries by run of the flow data for the total hydrocarbon run intervals (interval summary button will reposition spreadsheet). Moisture content and board weight data are on the "Define" worksheet and the original data are in the files named "Weights, CV-1.XLS".

HAPs

Within the file "HAPs, CV-1.xls", the summary page presents the data by run interval. The data is copied from the other pages to provide a concise summary.

The "Field Data" page is data transcribed from the field data sheets (copies of the sheets are included in Appendix 2 in PDF format) and includes the ambient pressure, lab temperature, flow rate through the impingers, and run start and stop times.

The "Laboratory Data" page contains results from the lab analysis for HAPs. These values come from the files "AQU, GC Sheet, CV-1.xls" and "ALD, GC Sheet, Cacade.xls" in the "Lab Data" directory. The GC retention times and peak areas and the GC calibrations are in these files.

On the "Impinger Calculations" page, the field data and laboratory data are used to give a dry gas flow rate through the impingers (columns J and K) and the mass of target compounds in the impingers (columns L to Q). Flow rates were adjusted to standard conditions in columns F and G.

ImpgrFlowstd_mL = ImpgrFlow * (273.16K / T_{meter}) / (P_{meter} / 101.33 kPa)

A dry gas flow rate is calculated in columns H and I

 $ImpgrFlow_{Dry_mL} = ImpgrFlow_{Std_mL} / (1 + MolHum)$

The average of the before and after gas flow measurements through the impingers (column J) is then converted to a mass basis in column K.

ImpgrFlow_{Dry_g} = MWair* ImpgrFlw_{Dry_mL}* P / (T * R)

Finally, the mass of each compound recovered from the impinger is calculated in columns L to S.

Mass_i = (Concentration_i) / (DenSolvent) * (Mass solvent)

The "Kiln Calculations" page uses a ratio of the dry gas flow through the kiln (calculated in the spreadsheets named "Kiln, CV-1.xls" and copied to column D) to the dry gas flow rate through the impinger to scale up the quantities and obtain the mass of each compound leaving the kiln (columns I to P).

On the "Emission" page, the amount of a HAP leaving the kiln is divided by the mass (in kg) or volume of wood (in mbf) to express the emissions on a per kg of wood (columns B-I) or per mbf basis (columns J-Q). Concentrations leaving the kiln are given in columns R to AG.

The "Quality Assurance" page presents information on the spikes, duplicates and blanks. For each spike a % recovery is calculated based on the mass of a HAP recovered divided by the amount added. The difference for each duplicate is calculated as a percentage from the difference between the impingers divided by the average mass collected after adjusting for impinger flow.

The remaining pages in "HAPs, CV-1.xls" are for graphing purposes.

9. Chain of custody information

Wood was collected by mill personnel and delivered to Oregon State by Columbia Vista. The wood was retained by Oregon State after delivery as documented in section 1. Field samples remained at Oregon State University.

10. Calibration documentation

	S	ensidyne	, LP	
Ce	II S/N: 1302015-S		Date: Fet	oruary 07, 2013
was	is is to certify that to calibrated using für prated by instrumen tandards and Techt ults:	n flowmeter MCS ts directly traceab	-102, which has le to the Nation	been
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	2004 1996 2001 2000	2005 2004 2002 2004	1 8 1 4	0.05 0.40 0.05 0.20
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Figure 14. Flow meter calibration.

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Figure 15. Certificates for calibration gases.

11. Anomalies

During part of total hydrocarbon run 4, room air was sampled rather than kiln air. This was from 18:02 hours to 24:03 hours. The values for this period were obtained by drawing a straight line between the good reading prior to the problem and the good readings after the problem. Kiln conditions were steady during this period and the data loss had no impact on the results. This was approximately 3% of the total sampling time.

During part of total hydrocarbon run 15, the calibration das was left on so that a high value for VOC was read. This was from 84:04 hours to 90:01 hours. The values for this period were obtained by drawing a straight line between the good reading prior to the problem and the good readings after the problem. Kiln conditions were steady during this period and the data loss had no impact on the results. This was approximately 3% of the total sampling time.

The impingers were left on for 2:15 instead of 1:30 during impinger run 2. This had no apparent impact on the results, although past experience suggests that method 105 recoveries go down as the interval length exceeds three hours.

12. Statement of validity

The statements in this report accurately represent the testing that occurred.

Michael R Milote

Michael R. Milota

Oregon Wood Innovation Center Department of Wood Science and Engineering 136 Richardson Hall Oregon State University Corvallis, OR 97331-5751 (541) 737-4210 V (541) 737-3385 F
Appendix 1. Detailed sampling procedures

Checks of kiln to record on log

Purpose: Ensure kiln is operating correctly

Clock time: Record from computer

Run time: Record from computer. Check the box if the computer screen being refreshed and time is advancing.

Box temperature: Read from plastic electrical enclosure on wall or on computer screen. The top and bottom numbers on controller should be similar and greater than the kiln temperature, 240°F.

Line temperature: Read from plastic electrical enclosure on wall or on computer screen. The top and bottom numbers on controller should be similar and greater than the box temperature, 245°F.

Valve temperature: Read from plastic electrical enclosure on wall or on computer screen. The top and bottom numbers on controller should be similar and greater than the line temperature, 250°F.

Dry-bulb temperature: Read from computer screen. Compare to paper graph to be sure it's correct. If it's not within a degree or two of the chart, check again in a few minutes. During startup (the first 3 or so hours), it may not be able to track. If it's too high, the heat valve should be closed, too low and the heat valve should be open. If it does not appear to be working correctly, call Mike.

Wet-bulb temperature: Read from computer screen. Compare to graph to be sure it's correct.

If the wet-bulb is too low, it means that the kiln atmosphere is too dry. Check the flow meters. If Flow1 is about 6 L/min (its lower limit), make sure that Flow2 and Flow3 are turned off. Flow2 records automatically. Enter any Flow3 change into the computer. Otherwise, call Mike.

If it's too high, then either the kiln atmosphere is too humid or the sock is not being wetted. If Flow 1 is near 200 L/min (its upper limit) add venting by opening Flow2 and/or Flow 3. Enter any Flow3 change into the computer. The maximum for Flow2 is 50 L/min, if it reads over this value for several readings, reduce it to about 45 L/min. Don't change Flow3 often, rather set it and leave it for several hours if possible. Keep the Flow 3 reading constant by small adjustments. As Flow1 decreases or Flow2 turned down, there is more pressure behind Flow3 and the flow increased. Check for water in the wet-bulb reservoir (push the float down and make sure it's getting water).

Check both Wet-bulb1 and Wet-bulb2 and make sure they are reading about the same. If they differ by more than 2°F, call Mike

If both wet-bulbs are reading the same as the dry-bulb, check the wet-bulb water.

If these procedures do not correct the wet-bulb temperature within 30 minutes, call Mike.

Chiller temperature: Read the chiller temperature. It should be about -1°C.

Flow 1: Read from computer. The value of Flow1 changes depending on the wetbulb. If Flow 1 is 6 L/min and the wet-bulb is too low, there's probably nothing we can do. If it's 200 L/min and the wet-bulb is too high, Flow2 and/or Flow3 can be opened. Flow2 and Flow3 should be adjusted so that Flow1 stays below 175 to 200 L/min.

Flow 2: Read from computer. The value of Flow2 is set by you. It will vary a little - as flow 1 goes down, flow 2 will go up. Do not set it to > 40 L/min if you think Flow1 is going to decrease or it will go off scale and not be read by the computer

Flow 3: Read from meter. The value of Flow3 is set by you. It will vary a little - as flow 1 goes down, flow 2 will go up. Be sure to clearly record this value and when you change it. Change it on the computer screen (click on it and type the new value).

Dilution flow: Read dilution flow meter. It should read the same setting as the red flag. Do not adjust. If significantly different, investigate.

Impinger flows: Read from rotometers. This should be about 250 to 500 cc/min.

Line vacuum: Read from the vacuum gauge. This should be about 20"Hg.

Total hydrocarbon analyzer

BACKGROUND INFORMATION

Get the dry- and wet-bulb temperatures from the kiln schedule or off the computer. Use the highest expected values for the next three to six hours.

Read absolute humidity off the psychrometric chart or table. Calculate or read from tables -

> Percent moisture = 100 / [1 + 1 / 1.61*AbHum] Target Dilution Ratio (TDR) = 15 / Percent Moisture

Event = the name of the drying cycle. Run = the number of the 3-hour interval. Operator, that's you. Date – use date VOC run will start if close to midnight

AMBIENT DATA

Read the laboratory temperature from the computer or thermometer.

ANALYZER CALIBRATION (BEFORE SIDE OF SHEET)

Set valves so that 1, 2 = OFF; 3=ON; 4=VENT. This allows gas to flow out of the vents from the calibration tanks and shuts off all other sources. Only calibration gas should go through the detector.

Open the zero gas tank valve set analyzer to range 3 zero valve on, others off set flow to 3 L/min using regulator on tank wait for a stable reading (about 30 to 60 seconds) use the zero dial (pot) on THA to get a zero reading read the analyzer read computer note pot setting Close valve on zero gas tank

Open span gas tank valve (may be 197 or 794 ppm gas) span valve on, others off set flow to 3 L/min using regulator on tank wait for a stable reading (about 30 to 60 seconds) use the span dial (pot) on THA to get a reading of 610ppm read the analyzer and record, eg, record 7.96 read computer (should read about 794) record pot setting Leave span tank valve open

Open mid gas tank valve (197 or 50 ppm gas) mid valve right on, others off set flow to 3 L/min using regulator on tank wait for a stable reading (about 30 to 60 seconds) read and record analyzer and computer (do not adjust pot settings) check for within tolerance switch analyzer to range 2 read analyzer and computer check for within tolerance switch analyzer back to range 3 Turn off mid gas tank valve

SET DILUTION FLOW BEFORE RUN (BEFORE SIDE OF SHEET)

Set valves so that 1, 2, 3 = OFF; 4=meter. This allows gas to flow only from the meter to the detector.

Use the Gilibrator to take 4 readings of the total flow rate (TFR). This is the total flow drawn by the analyzer and should be about 1.6 L/min Make sure the average does not include any "bad" readings Record the average in mL/min; It should be 1500-1600 mL/min Write the Run # and "Pre-TFR" on the Gilibrator printout.

Calculate the next two values -Target dilution flow rate (TDFR) is the TFR x (1 - DR) Target sample flow rate (TSFR) is the TFR x DR Check that the sum of these is the Total Flow Rate

Set dilution flow Set red pointer to desired dilution flow Slowly open lower valve on dilution flow meter (1=ON) Use upper valve on dilution flow meter to adjust flow Do not adjust this meter after this point Read the meter that you just set and record the value in SCFH Calculate and record L/min

Use the Gilibrator to take 4 readings of the sample flow rate (SFR). This is the flow through the analyzer after dilution is set. It will vary, depending on the dilution setting.

Make sure the average does not include any "bad" readings Record the average in mL/min

Write "Pre-SFR" on the Gilibrator printout.

CHECK DILUTION FLOW BEFORE RUN (BEFORE SIDE OF SHEET)

Set valves so that 1, 3 = ON; 2=OFF; 4=VENT. This allows gas to flow out of the vent from the calibration tank and shuts off all other sources. Calibration gas and dilution air will go through the detector.

Open span gas tank valve (should already be open) span panel valve right (on), others down (off) set flow to 3 L/min using regulator on tank set analyzer to range 3 wait for a stable reading (about 30 to 60 seconds), record turn off all calibration gas tank valves all calibration gas panel valves off

All tank valves off

Calculate the dilution ratio based on gas flow by dividing the Sample Flow Rate by the Total Flow Rate. DR = Absolute value of [100*(DR Span - DR Flow)/DR Flow]

Calculate the dilution ratio based on span gas by dividing the diluted span by the undiluted span.

If the dilution ratio calculated from the span gas and the dilution ration calculated from the flow do not agree within 5% - DO NOT PROCEED****. Check the calculations, then redo the measurements.

**** check calculations, check that values for ppm and flows make sense, remeasure everything. If it still does not agree, call Mike

START RUN (BOTTOM OF BEFORE SIDE OF SHEET)

Set valve so that 1, 2, 5 = on; 3, 4=off; all calibration tank valves off

Record the start time. Use the computer clock or stopwatch time.

Make sure analyzer is on appropriate range, usually range 3, to keep THC reading on computer between 60 and 600.

Monitor system, as needed. Record system condition at least hourly.

End time should be no more than 3-6 hours from start time.

POST-SAMPLE PROCEDURE - AT END OF RUN (AFTER SIDE OF SHEET)

Record your name as the operator.

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Event = the drying cycle. Run = number of the 3-hour interval. Operator, that's you.

AMBIENT DATA

Read the laboratory temperature from the thermometer.

Fill out appropriate information on Pre-sample side of data sheet for next run. This will save time in between runs.

END TIME

Record computer time.

DO NOT adjust dilution gas or analyzer pots until the instructions tell you to.

CHECK DILUTION FLOW AFTER RUN (AFTER SIDE OF SHEET)

Measure diluted span gas: Set valves so that 1, 3 = on; 2=off; 4=vent. This allows gas to flow out of the vent from the calibration tank and shuts off all other sources. Calibration gas and dilution air will go through the detector.

Open span gas tank valve

span panel valve ON, others OFF set flow to 3 L/min using regulator on tank set analyzer to range 3 wait for a stable reading (about 30 -60 seconds) record

Sample flow rate: Set values so that 1=on; 2, 3 = off; 4=meter. This allows gas to flow only from the meter and the dilution to the detector.

Use the Gilibrator to take 4 readings of the sample flow rate (SFR). This is the flow through the analyzer with dilution on. Make sure the average does not include any "bad" readings Record the average in L/min Write Run # and "Post-SFR" on the Gilibrator printout.

Read dilution flow meter To calculate the L/min, divide scfh by 2.12 Turn off dilution flow meter using valve 1 (lower dilution valve)

Total flow rate. Set valves so that 1, 2, 3 = off; 4=meter. This allows gas to flow only from the meter to the detector.

Use the Gilibrator to take 4 readings of the total flow rate (TFR). This is the total flow drawn by the analyzer and should be about 1.6 L/min Make sure the average does not include any "bad" readings Record the average Write Run # and "Post-TFR" on the Gilibrator printout.

Calculate the dilution ratio based on gas flow by dividing the Sample Flow Rate by the Total Flow Rate.

CHECK CALIBRATION OF ANALYZER (AFTER SIDE OF SHEET)

Set valves so that 1, 2 = off; 3=on; 4=vent. This allows gas to flow out of the vents from the calibration tanks and shuts off all other sources. Only calibration gas should go through the detector.

Span gas tank valve should be open

span panel valve ON, others down OFF set flow to 3 L/min using regulator on tank set analyzer to range 3 wait for a stable reading (about 30 -60 seconds) read analyzer (do not adjust pot settings), record, for example, 7.94 as 794 read computer (should read about the same) note pot setting check for within tolerance

Open mid gas tank valve

mid panel valve = ON, others OFF set flow to 3 L/min using regulator on tank set analyzer to range 3 wait for a stable reading (about 30 -60 seconds) read analyzer (do not adjust pot settings), record, for example, 1.97 as 197 read computer (should read same as analyzer) check for within tolerance

Open the zero gas tank valve

zero panel valve = ON, others OFF set flow to 3 L/min using regulator on tank wait for a stable reading (about 30 -60 seconds) read analyzer (do not adjust pot settings) read computer note pot setting

Close all tank valves if charge is ending

Calculate the dilution ratio based on gas concentration by dividing the Diluted span by the Span

Calculate % difference in the two dilution ratios as 100 * {Absolute Value (DRSpan-DRFlow)} / DRFlow

Record the time now as the end time for check.

Start Pre-Sample procedure for next run.

HAP 105 Collection

BACKGROUND DATA

Begin about 15 minutes before run should start Operator, that's you. Date, today or tomorrow if sample will start after midnight Event = Kiln Charge Run = sequence of M/F measurement (1-A, or 5-C, etc)

PRE RUN DATA

Call 9-541-754-0081 and get altimeter setting.

IMPINGER WEIGHTS

Verify that the impinger weights match the prerecorded weights on the data sheet.

Put 15 mL of BHA solution in impinger #1. Put 10 mL of BHA solution in impinger #2. Impinger #3 is not filled. It is for overflow.

Reweigh the impingers with the BHA solution. Place BHA stock back into cooler Install impingers and lower into chiller

LEAK CHECK

Read the laboratory temperature. Close valve to sample probe. Turn on pump (it may already be on) Evacuate to 15 to 18 " Hg, record Close valve that is near pump Note pressure and start timer Allowable pressure change is 1" Hg in 2 minutes, if it is more than this, find the source of the leak. Record change. Slowly open valve near probe tip so that pressure is slowly relieved. Completely open valve near probe tip Open valve near pump

SAMPLE FLOW RATE

Attach probe tip to Gilibrator Take 4 readings Make sure all readings in average are "good" readings Record the average

Columbia Vista, Douglas-fir

START TIME

Put probe into kiln (or turn valve to sample kiln) and record time. Check meters to make sure gas is flowing

FLOW READINGS DURING TEST

Note flow meter reading at intervals of at least 20-30 minutes Run test for 1:30 or less if impingers fill

POST RUN DATA

Begin about 10 minutes before run should end Label a sample bottle with the Event and Run numbers and record the weight. Call 9-541-754-0081 and get altimeter setting.

END TIME

Remove probe (or turn valve to meter setting) from kiln Record time

SAMPLE FLOW RATE

Rinse probe with 5 mL of DI water Read the laboratory. Attach probe tip to Gilibrator Take 5 readings Make sure all readings in average are "good" readings Record the average

IMPINGER WEIGHTS

Lift impingers from chiller, take to scale, and place onto rack Dry the outside of the impingers Remove U tubes connecting the impingers together Weigh sample bottle with lid Weigh the impingers (without stoppers) with the catch and record Transfer the impinger contents to the sample bottle Weigh the sample bottle with lid and record Rinse impingers (last to first) with 10 mL DIW (save in the sample bottle) Weigh the sample bottle with lid and record Rinser impingers (last to first) with 5 mL hexane (save in the sample bottle) Weigh the sample bottle with lid and record Rinser impingers (last to first) with 5 mL hexane (save in the sample bottle) Weigh the sample bottle into cold storage Record the volume of any liquids lost during this procedure. Wash glassware with phosphate-free detergent and set out to dry.



Uri ____ Randy Caul Wess _ Clint John Vannessa Jorry Grian Duane Allison Chip Traci Tina, File

Appendix 2. Electronic copy of data and calculations



